



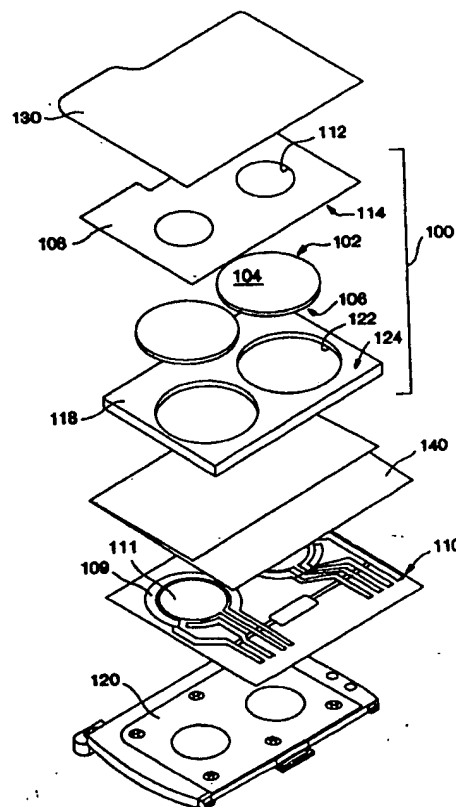
INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶ : A61N 1/30	A1	(11) International Publication Number: WO 99/58190 (43) International Publication Date: 18 November 1999 (18.11.99)
(21) International Application Number: PCT/US99/10378 (22) International Filing Date: 11 May 1999 (11.05.99) (30) Priority Data: 60/085,345 13 May 1998 (13.05.98) US (71) Applicant: CYGNUS, INC. [US/US]; 400 Penobscot Drive, Redwood City, CA 94063 (US). (72) Inventors: CONN, Thomas, E.; 257 Kingsley Avenue, Palo Alto, CA 94301 (US). FORD, Russell; 974 Jackson Street, San Francisco, CA 94133 (US). SONI, Pravin, L.; 1320 Bedford Avenue, Sunnyvale, CA 94087 (US). TIERNEY, Michael, J.; 368 North Sixth Street, San Jose, CA 95112 (US). VIJAYAKUMAR, Prema; 43493 Southerland Way, Fremont, CA 94539 (US). (74) Agents: McCLUNG, Barbara; Cygnus, Inc., 400 Penobscot Drive, Redwood City, CA 94063 (US) et al.		(81) Designated States: CA, JP, KR, European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE). Published <i>With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i>

(54) Title: COLLECTION ASSEMBLIES FOR TRANSDERMAL SAMPLING SYSTEM

(57) Abstract

The invention relates generally to consumable components of a device used for continually or continuously measuring the concentration of target chemical analytes present in a biological system. More particularly, the invention relates to collection assemblies (100), laminate structures, and autosensor assemblies, which are used in connection with a transdermal sampling device. In one aspect, the invention includes autosensor assemblies which include laminate structures (130, 108, 118, 140), electrode assemblies (109, 110, 111), and support trays (120). One important application of the invention involves an autosensor assembly for use in a blood glucose monitoring device.



FOR THE PURPOSES OF INFORMATION ONLY

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AL	Albania	ES	Spain	LS	Lesotho	SI	Slovenia
AM	Armenia	FI	Finland	LT	Lithuania	SK	Slovakia
AT	Austria	FR	France	LU	Luxembourg	SN	Senegal
AU	Australia	GA	Gabon	LV	Latvia	SZ	Swaziland
AZ	Azerbaijan	GB	United Kingdom	MC	Monaco	TD	Chad
BA	Bosnia and Herzegovina	GE	Georgia	MD	Republic of Moldova	TG	Togo
BB	Barbados	GH	Ghana	MG	Madagascar	TJ	Tajikistan
BE	Belgium	GN	Guinea	MK	The former Yugoslav Republic of Macedonia	TM	Turkmenistan
BF	Burkina Faso	GR	Greece			TR	Turkey
BG	Bulgaria	HU	Hungary	ML	Mali	TT	Trinidad and Tobago
BJ	Benin	IE	Ireland	MN	Mongolia	UA	Ukraine
BR	Brazil	IL	Israel	MR	Mauritania	UG	Uganda
BY	Belarus	IS	Iceland	MW	Malawi	US	United States of America
CA	Canada	IT	Italy	MX	Mexico	UZ	Uzbekistan
CF	Central African Republic	JP	Japan	NE	Niger	VN	Viet Nam
CG	Congo	KE	Kenya	NL	Netherlands	YU	Yugoslavia
CH	Switzerland	KG	Kyrgyzstan	NO	Norway	ZW	Zimbabwe
CI	Côte d'Ivoire	KP	Democratic People's Republic of Korea	NZ	New Zealand		
CM	Cameroon			PL	Poland		
CN	China	KR	Republic of Korea	PT	Portugal		
CU	Cuba	KZ	Kazakstan	RO	Romania		
CZ	Czech Republic	LC	Saint Lucia	RU	Russian Federation		
DE	Germany	LI	Liechtenstein	SD	Sudan		
DK	Denmark	LK	Sri Lanka	SE	Sweden		
EE	Estonia	LR	Liberia	SG	Singapore		

COLLECTION ASSEMBLIES FOR TRANSDERMAL SAMPLING SYSTEM

TECHNICAL FIELD OF THE INVENTION

5 Novel laminate structures, collection assemblies, and autosensor assemblies for use in a sampling device are described. The invention relates generally to consumable components of a device used for continually or continuously measuring the concentration of target
10 chemical analytes present in a biological system. The laminates, collection assemblies, and autosensor assemblies are used in a transdermal sampling device that is placed in operative contact with a skin or mucosal surface of a biological system to obtain a
15 chemical signal associated with an analyte of interest.

BACKGROUND OF THE INVENTION

 A number of diagnostic tests are routinely performed on humans to evaluate the amount or existence
20 of substances present in blood or other body fluids. These diagnostic tests typically rely on physiological fluid samples removed from a subject, either using a syringe or by pricking the skin. One particular diagnostic test entails self-monitoring of blood glucose
25 levels by diabetics.

 Diabetes is a major health concern, and treatment of the more severe form of the condition, Type I (insulin-dependent) diabetes, requires one or more insulin injections per day. Insulin controls
30 utilization of glucose or sugar in the blood and prevents hyperglycemia which, if left uncorrected, can

lead to ketosis. On the other hand, improper administration of insulin therapy can result in hypoglycemic episodes, which can cause coma and death. Hyperglycemia in diabetics has been correlated with several long-term effects of diabetes, such as heart disease, atherosclerosis, blindness, stroke, hypertension and kidney failure.

The value of frequent monitoring of blood glucose as a means to avoid or at least minimize the complications of Type I diabetes is well established. Patients with Type II (non-insulin-dependent) diabetes can also benefit from blood glucose monitoring in the control of their condition by way of diet and exercise.

Conventional blood glucose monitoring methods generally require the drawing of a blood sample (e.g., by finger prick) for each test, and a determination of the glucose level using an instrument that reads glucose concentrations by electrochemical or colorimetric methods. Type I diabetics must obtain several finger prick blood glucose measurements each day in order to maintain tight glycemic control. However, the pain and inconvenience associated with this blood sampling, along with the fear of hypoglycemia, has led to poor patient compliance, despite strong evidence that tight control dramatically reduces long-term diabetic complications. In fact, these considerations can often lead to an abatement of the monitoring process by the diabetic. See, e.g., The Diabetes Control and Complications Trial Research Group (1993) *New Engl. J. Med.* 329:977-1036.

Recently, various methods for determining the concentration of blood analytes without drawing blood have been developed. For example, U.S. Patent No.

5,267,152 to Yang et al. describes a noninvasive technique of measuring blood glucose concentration using near-IR radiation diffuse-reflection laser spectroscopy. Similar near-IR spectrometric devices are also described
5 in U.S. Patent No. 5,086,229 to Rosenthal et al. and U.S. Patent No. 4,975,581 to Robinson et al.

U.S. Patent Nos. 5,139,023 to Stanley et al., and 5,443,080 to D'Angelo et al. describe transdermal blood glucose monitoring devices that rely on a permeability
10 enhancer (e.g., a bile salt) to facilitate transdermal movement of glucose along a concentration gradient established between interstitial fluid and a receiving medium. U.S. Patent No. 5,036,861 to Sembrowich describes a passive glucose monitor that collects
15 perspiration through a skin patch, where a cholinergic agent is used to stimulate perspiration secretion from the eccrine sweat gland. Similar perspiration collection devices are described in U.S. Patent No. 5,076,273 to Schoendorfer and U.S. Patent No. 5,140,985
20 to Schroeder.

In addition, U.S. Patent No. 5,279,543 to Glikfeld et al. describes the use of iontophoresis to noninvasively sample a substance through skin into a receptacle on the skin surface. Glikfeld teaches that
25 this sampling procedure can be coupled with a glucose-specific biosensor or glucose-specific electrodes in order to monitor blood glucose. Finally, International Publication No. WO 96/00110, published 4 January 1996, describes an iontophoretic apparatus for transdermal
30 monitoring of a target substance, wherein an iontophoretic electrode is used to move an analyte into a collection reservoir and a biosensor is used to detect

the target analyte present in the reservoir. Finally, International Publication No. WO 96/00110 to Tamada describes an iontophoretic apparatus for transdermal monitoring of a target substance, where an iontophoretic
5 electrode is used to move an analyte into a collection reservoir and a biosensor is used to detect the target analyte present in the reservoir.

SUMMARY OF THE INVENTION

10 The present invention relates generally to collection assembly, laminates and autosensor assemblies for use in a sampling device. More particularly, the present collection assembly, laminates and autosensor
15 assemblies are used in a transdermal sampling device that is placed in operative contact with a skin or mucosal surface of the biological system to obtain a chemical signal associated with an analyte of interest. The sampling device transdermally extracts the analyte from the biological system using, for example, an
20 iontophoretic sampling technique. The transdermal sampling device can be maintained in operative contact with the skin or mucosal surface of the biological system to provide, for example, continual or continuous analyte measurement.

25 The analyte can be any specific substance or component that one is desirous of detecting and/or measuring in a chemical, physical, enzymatic, or optical analysis. The analyte can be any specific substance or component that one is desirous of detecting and/or
30 measuring in a chemical, physical, enzymatic, or optical analysis. Such analytes include, but are not limited

to, amino acids, enzyme substrates or products indicating a disease state or condition, other markers of disease states or conditions, drugs of abuse, therapeutic and/or pharmacologic agents (e.g., theophylline, anti-HIV drugs, lithium, anti-epileptic drugs, cyclosporin, chemotherapeutics), electrolytes, physiological analytes of interest (e.g., urate/uric acid, carbonate, calcium, potassium, sodium, chloride, bicarbonate (CO₂), glucose, urea (blood urea nitrogen), lactate/lactic acid, hydroxybutyrate, cholesterol, triglycerides, creatine, creatinine, insulin, hematocrit, and hemoglobin), blood gases (carbon dioxide, oxygen, pH), lipids, heavy metals (e.g., lead, copper), and the like. In preferred embodiments, the analyte is a physiological analyte of interest, for example glucose, or a chemical that has a physiological action, for example a drug or pharmacological agent.

Thus, one embodiment of the invention provides a tri-layer collection assembly for use in a transdermal sampling device. The collection assembly is formed from a series of functional layers including: (1) a first surface layer that is comprised of a substantially planar material that has an opening which extends therethrough; (2) a second surface layer that is also comprised of a substantially planar material and has an opening therein; and (3) an intervening layer that is positioned between the first and second surface layers, wherein the intervening layer is comprised of an ionically conductive material. The first and second surface layers overlap the intervening layer at

corresponding positions, and contact each other at their corresponding overlaps, such overlaps can be used to form a laminate structure. The openings in the first and second surface layers are axially aligned to provide
5 a flow path through the laminate (i.e., a flow path that extends between the two surfaces and passes through the intervening layer). The overhangs provided by the mask and retaining layers are generally contacted with each other to sandwich the collection insert therebetween and
10 form the assembly.

It is a related object of the invention to provide an autosensor assembly for use in a transdermal sampling device, wherein the assembly comprises the three functional layers of the above-described collection
15 assembly or laminate, an electrode assembly, and, typically, a support tray.

It is a further object of the invention to provide a two layer collection assembly or laminate for use in a transdermal sampling device. The collection assembly is
20 formed from two functional layers including: (1) a surface layer that is comprised of a substantially planar material that has an opening which extends therethrough; and (2) a second layer. The second layer is formed from the combination of a gasket and a
25 collection insert. The gasket is comprised of a substantially planar material having a top face, a bottom face, and an opening extending between the top and bottom faces. The top face of the gasket is attached to the bottom face of the surface layer, and
30 the opening in the gasket is axially aligned with the

opening in the surface layer to provide a flow path through the laminate. The collection insert is arranged within and substantially fills the opening in the gasket such that the collection insert is aligned with the opening in the surface layer and rests against or is otherwise attached to a portion of the surface layer.

It is a related object of the invention to provide an autosensor assembly for use in a transdermal sampling device, wherein the assembly comprises the two layers of the above-described collection assembly or laminate, an electrode assembly with which the collection assembly is functionally aligned, and, typically, a support tray.

Thus, in one embodiment, the invention relates to a collection assembly for use in a iontophoretic sampling device useful to monitor a selected analyte or derivatives thereof present in a biological system. The collection assembly comprises:

a) a collection insert layer comprised of an ionically conductive material having first and second portions, each portion having first and second surfaces,

b) a mask layer comprised of a material that is substantially impermeable to the selected analyte or derivatives thereof, wherein the mask layer (i) has inner and outer faces and said outer face provides contact with said biological system and the inner face is positioned in facing relation with the first surface of each collection insert, (ii) defines first and second openings that are aligned with the first and second portions of the collection insert layer, (iii) each opening exposes at least a portion of the first surface

of the collection insert layer, and (iv) has a border which extends beyond the first surface of each portion of the collection insert layer to provide an overhang; and

5 (c) a retaining layer having (i) inner and outer faces wherein the inner face is positioned in facing relation with the second surface of each collection insert, (ii) defines first and second openings that are aligned with the first and second portions of the
10 collection insert layer, (iii) each opening exposes at least a portion of the second surface of the collection insert layer, and (iv) has a border which extends beyond the first surface of each portion of the collection insert layer to provide an overhang.

15 In certain embodiments, the collection insert layer further comprises a gasket layer and the gasket layer is between the mask layer and the retaining layer.

In additional embodiments, the subject invention is directed to a laminate comprising a collection assembly,
20 as described above, as well as a sealed package containing the laminate.

In still another embodiment, the invention is directed to an autosensor assembly for use in a iontophoretic sampling device useful to monitor an
25 analyte present in a biological system. The autosensor assembly comprises:

(I) a collection assembly which comprises,
a) a collection insert layer comprised of an ionically conductive material having first and second
30 portions, each portion having first and second surfaces,

b) a mask layer comprised of a substantially planar material that is substantially impermeable to the selected analyte or derivatives thereof, wherein the mask layer (i) has inner and outer faces and the outer
5 face provides contact with the biological system and the inner face is positioned in facing relation with the first surface of each collection insert, (ii) defines first and second openings that are aligned with the first and second portions of the collection insert
10 layer, (iii) each opening exposes at least a portion of the first surface of the collection insert layer, and (iv) has a border which extends beyond the first surface of each portion of the collection insert layer to provide an overhang;

15 (c) a retaining layer having (i) inner and outer faces wherein the inner face is positioned in facing relation with the second surface of each collection insert, (ii) defines first and second openings that are aligned with the first and second portions of the
20 collection insert layer, (iii) each opening exposes at least a portion of the second surface of the collection insert layer, and (iv) has a border which extends beyond the first surface of each portion of the collection insert layer to provide an overhang; and

25 (d) where the first and second openings in the mask layer are positioned in the collection assembly such that they are aligned with the first and second openings in the retaining layer and thereby define a plurality of flow paths through said collection assembly;

(II) an electrode assembly having an inner and outer face, the inner face comprising first and second bimodal electrodes, wherein the first and second bimodal electrodes are aligned with the first and second openings in the retaining layer of the collection assembly; and

(III) a support tray that contacts the outer face of the electrode assembly.

In alternative embodiments, the autosensor assembly further comprises a first removable liner attached to the outer face of the retaining layer, and/or a second removable liner attached to the outer face of the mask layer. In addition, a plowfold liner can be used, for example, between the electrode surfaces and the collection inserts.

In further embodiments, the invention is directed to a sealed package containing the autosensor assembly described above. The sealed package may also contain a hydrating insert.

In yet another embodiment, the invention is directed to a collection assembly for use in a iontophoretic sampling device useful to monitor a selected analyte, or derivatives thereof, present in a biological system. The collection assembly comprises:

a) a mask layer comprised of a substantially planar material that is substantially impermeable to the selected analyte or derivatives thereof, where the mask layer has inner and outer faces and the outer face provides contact with the biological system;

b) a collection insert layer comprised of an ionically conductive material having first and second surfaces, and

c) the mask and collection insert layers are
5 configured such that (i) at least a portion of the collection insert is exposed to provide contact with the biological system, and (ii) flow of the analyte through the first surface of the collection insert layer from the biological system is prevented by the mask layer for
10 any portion of the first surface of the collection insert layer that is in contact with the inner face of the mask layer.

In another embodiment, the invention is directed to an autosensor assembly comprising

15 (a) the collection assembly above,
(b) an electrode assembly having an inner face comprising an electrode and an outer face, where the inner face of the electrode assembly and the collection assembly are aligned to define a plurality of flow paths
20 through the collection assembly, and
(c) a support tray that contacts the outer face of the electrode assembly.

Additional objects, advantages and novel features
25 of the invention will be set forth in part in the description which follows, and in part will become apparent to those skilled in the art upon examination of the following, or may be learned by practice of the invention.

30

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1A depicts a top plan view of an iontophoretic collection reservoir and electrode assembly for use in a transdermal sampling device.

5 Figure 1B depicts the side view of the iontophoretic collection reservoir and electrode assembly shown in Figure 1A.

10 Figure 2 is a pictorial representation of an iontophoretic sampling device which includes the iontophoretic collection reservoir and electrode assembly of Figures 1A and 1B.

Figure 3 depicts an exploded view of one embodiment of a collection assembly and autosensor constructed according to the present invention.

15 Figure 4 depicts an exploded view of another embodiment of a collection assembly and autosensor constructed according to the present invention.

20 Figure 5 depicts an exploded view of a still further embodiment of a collection assembly and autosensor constructed according to the present invention.

Figure 6 is a representation of one embodiment of a bimodal electrode design. The figure presents an overhead and schematic view of the electrode assembly
25 633. In the figure, the bimodal electrode is shown at 630 and can be, for example, a Ag/AgCl iontophoretic/counter electrode. The sensing or working electrode (made from, for example, platinum) is shown at 631. The reference electrode is shown at 632 and can
30 be, for example, a Ag/AgCl electrode. The components are mounted on a suitable nonconductive substrate 634, for example, plastic or ceramic. The conductive leads

637 leading to the connection pad 635 are covered by a second nonconductive piece 636 of similar or different material. In this example of such an electrode the working electrode area is approximately 1.35 cm^2 . The dashed line in Figure 6 represents the plane of the cross-sectional schematic view presented in Figure 7.

Figure 7 is a representation of a cross-sectional schematic view of the bimodal electrodes as they may be used in conjunction with a reference electrode and a hydrogel pad. In the figure, the components are as follows: bimodal electrodes 740 and 741; sensing electrodes 742 and 743; reference electrodes 744 and 745; a substrate 746; and hydrogel pads 747 and 748.

Figures 8A through 8H show general schematic diagrams for the components of one embodiment of an autosensor of the present invention. The general shape and dimensions of the tray are indicated in Figure 8A. General shape and dimensions of the electrode assembly are indicated in Figure 8B. A tri-layer laminate including a mask layer, having the general shape and dimensions shown in Figure 8C, collection inserts, having the general shape and dimensions shown in Figure 8D, and a retaining layer, having the general shape and dimensions shown in Figure 8E. Further, Figure 8F shows the general shape and dimensions of the liner for contacting the mask and collection inserts (i.e., a "patient liner"). Figure 8G shows the general shape and dimensions of the second liner (i.e., a "plow-fold" liner), for contacting the retaining layer and collection inserts. Figure 8H shows a composite figure of autosensor components in their proper order of stacking/assembly.

Figures 9A through 9G show general schematic diagrams for the components of another embodiment of an autosensor of the present invention. The general shape and dimensions of the tray are indicated in Figure 9A.

5 General shape and dimensions of the electrode assembly are indicated in Figure 9B. A mask layer is shown, having the general shape and dimensions shown in Figure 9C. The general shape and dimensions of collection inserts are shown in Figure 9D. Figure 9E shows the

10 general shape and dimensions of the second liner (i.e., a "plow-fold" liner), for contacting the gel retaining layer and the collection inserts, typically eliminating contact between the collection inserts and the iontophoretic/counter electrodes and reference electrode

15 prior to removal. Further, Figure 9F shows the general shape and dimensions of the liner for contacting the mask layer and collection inserts (i.e., a "patient liner"). Figure 9G shows a composite figure of autosensor components in their proper order of

20 stacking/assembly.

DETAILED DESCRIPTION OF THE INVENTION

Before describing the present invention in detail, it is to be understood that this invention is not

25 limited to particular compositions or biological systems as such may, of course, vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to be limiting.

30 It must be noted that, as used in this specification and the appended claims, the singular forms "a", "an" and "the" include plural referents

unless the content clearly dictates otherwise. Thus, for example, reference to "a collection insert" includes two or more such inserts, reference to "an analyte" includes a mixture of two or more such analytes, reference to "an electrochemically active species" includes two or more such species, and the like.

Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which the invention pertains. Although any methods and materials similar or equivalent to those described herein can be used in the practice of the present invention, the preferred materials and methods are described herein.

In describing and claiming the present invention, the following terminology will be used in accordance with the definitions set out below.

1. DEFINITIONS

The terms "analyte" and "target analyte" are used herein to denote any physiological analyte of interest that is a specific substance or component that is being detected and/or measured in a chemical, physical, enzymatic, or optical analysis. A detectable signal (e.g., a chemical signal or electrochemical signal) can be obtained, either directly or indirectly, from such an analyte or derivatives thereof. Furthermore, the terms "analyte" and "substance" are used interchangeably herein, and are intended to have the same meaning, and thus encompass any substance of interest. In preferred embodiments, the analyte is a physiological analyte of interest, for example, glucose, or a chemical that has a

physiological action, for example, a drug or pharmacological agent.

A "sampling device" or "sampling system" refers to any device for obtaining a sample from a biological system for the purpose of determining the concentration of an analyte of interest. As used herein, the term "sampling" means invasive, minimally invasive or non-invasive extraction of a substance from the biological system, generally across a membrane such as skin or mucosa. The membrane can be natural or artificial, and can be of plant or animal nature, such as natural or artificial skin, blood vessel tissue, intestinal tissue, and the like. Typically, the sampling means are in operative contact with a "reservoir," or "collection reservoir," wherein the sampling means is used for extracting the analyte from the biological system into the reservoir to obtain the analyte in the reservoir. A "biological system" includes both living and artificially maintained systems. Examples of minimally invasive and noninvasive sampling techniques include iontophoresis, sonophoresis, suction, electroporation, thermal poration, passive diffusion, microfine (miniature) lances or cannulas, subcutaneous implants or insertions, and laser devices. Sonophoresis uses ultrasound to increase the permeability of the skin (see, e.g., Menon et al. (1994) *Skin Pharmacology* 7:130-139). Suitable sonophoresis sampling systems are described in International Publication No. WO 91/12772, published 5 September 1991. Passive diffusion sampling devices are described, for example, in International Publication Nos.: WO 97/38126 (published 16 October 1997); WO 97/42888, WO 97/42886, WO 97/42885, and WO

97/42882 (all published 20 November 1997); and WO
97/43962 (published 27 November 1997). Laser devices
use a small laser beam to burn a hole through the upper
layer of the patient's skin (see, e.g., Jacques et al.
5 (1978) *J. Invest. Dermatology* 88:88-93). Examples of
invasive sampling techniques include traditional needle
and syringe or vacuum sample tube devices.

A "housing" for the sampling system can further
include suitable electronics (e.g., microprocessor,
10 memory, display and other circuit components) and power
sources for operating the sampling system in an
automatic fashion.

A "monitoring system," as used herein, refers to a
system useful for continually or continuously measuring
15 a physiological analyte present in a biological system.
Such a system typically includes, but is not limited to,
sampling means, sensing means, and a microprocessor
means in operative communication with the sampling means
and the sensing means.

20 The term "artificial," as used herein, refers to an
aggregation of cells of monolayer thickness or greater
which are grown or cultured *in vivo* or *in vitro*, and
which function as a tissue of an organism but are not
actually derived, or excised, from a pre-existing source
25 or host.

The term "subject" encompasses any warm-blooded
animal, particularly including a member of the class
Mammalia such as, without limitation, humans and
nonhuman primates such as chimpanzees and other apes and
30 monkey species; farm animals such as cattle, sheep,
pigs, goats and horses; domestic mammals such as dogs
and cats; laboratory animals including rodents such as

mice, rats and guinea pigs, and the like. The term does not denote a particular age or sex. Thus, adult and newborn subjects, as well as fetuses, whether male or female, are intended to be covered.

5 As used herein, the term "continual measurement" intends a series of two or more measurements obtained from a particular biological system, which measurements are obtained using a single device maintained in operative contact with the biological system over the
10 time period in which the series of measurements is obtained. The term thus includes continuous measurements.

 The term "transdermal," as used herein, includes both transdermal and transmucosal techniques, i.e.,
15 extraction of a target analyte across skin or mucosal tissue. Aspects of the invention which are described herein in the context of "transdermal," unless otherwise specified, are meant to apply to both transdermal and transmucosal techniques.

20 The term "transdermal extraction," or "transdermally extracted" intends any noninvasive, or at least minimally invasive sampling method, which entails extracting and/or transporting an analyte from beneath a tissue surface across skin or mucosal tissue. The term
25 thus includes extraction of an analyte using iontophoresis (reverse iontophoresis), electroosmosis, sonophoresis, microdialysis, suction, and passive diffusion. These methods can, of course, be coupled with application of skin penetration enhancers or skin
30 permeability enhancing technique such as tape stripping or pricking with micro-needles. The term "transdermally extracted" also encompasses extraction techniques which

employ thermal poration, electroporation, microfine lances, microfine canulas, subcutaneous implants or insertions, and the like.

The term "iontophoresis" intends a method for
5 transporting substances across tissue by way of an application of electrical energy to the tissue. In conventional iontophoresis, a reservoir is provided at the tissue surface to serve as a container of material to be transported. Iontophoresis can be carried out
10 using standard methods known to those of skill in the art, for example, by establishing an electrical potential using a direct current (DC) between fixed anode and cathode "iontophoretic electrodes," alternating a direct current between anode and cathode
15 iontophoretic electrodes, or using a more complex waveform such as applying a current with alternating polarity (AP) between iontophoretic electrodes (so that each electrode is alternately an anode or a cathode).

The term "reverse iontophoresis" refers to the
20 movement of a substance from a biological fluid across a membrane by way of an applied electric potential or current. In reverse iontophoresis, a reservoir is provided at the tissue surface to receive the extracted material.

25 "Electroosmosis" refers to the movement of a substance through a membrane by way of an electric field-induced convective flow. The terms iontophoresis, reverse iontophoresis, and electroosmosis, will be used interchangeably herein to refer to movement of any
30 ionically charged or uncharged substance across a membrane (e.g., an epithelial membrane) upon application

of an electric potential to the membrane through an ionically conductive medium.

The term "sensing device," "sensing means," or "biosensor device" encompasses any device that can be used to measure the concentration of an analyte, or derivative thereof, of interest. Preferred sensing devices for detecting blood analytes generally include electrochemical devices and chemical devices. Examples of electrochemical devices include the Clark electrode system (see, e.g., Updike, et al., (1967) *Nature* 214:986-988), and other amperometric, coulometric, or potentiometric electrochemical devices. Examples of chemical devices include conventional enzyme-based reactions as used in the Lifescan® glucose monitor (Johnson and Johnson, New Brunswick, NJ) (see, e.g., U.S. Patent 4,935,346 to Phillips, et al.).

A "biosensor" or "biosensor device" includes, but is not limited to, a "sensor element" which includes, but is not limited to, a "biosensor electrode" or "sensing electrode" or "working electrode" which refers to the electrode that is monitored to determine the amount of electrical signal at a point in time or over a given time period, which signal is then correlated with the concentration of a chemical compound. The sensing electrode comprises a reactive surface which converts the analyte, or a derivative thereof, to electrical signal. The reactive surface can be comprised of any electrically conductive material such as, but not limited to, platinum-group metals (including, platinum, palladium, rhodium, ruthenium, osmium, and iridium), nickel, copper, silver, and carbon, as well as, oxides, dioxides, combinations or alloys thereof. Some

catalytic materials, membranes, and fabrication technologies suitable for the construction of amperometric biosensors were described by Newman, J.D., et al. (*Analytical Chemistry* 67(24), 4594-4599, 1995).

5 The "sensor element" can include components in addition to a biosensor electrode, for example, it can include a "reference electrode," and a "counter electrode." The term "reference electrode" is used herein to mean an electrode that provides a reference
10 potential, e.g., a potential can be established between a reference electrode and a working electrode. The term "counter electrode" is used herein to mean an electrode in an electrochemical circuit which acts as a current source or sink to complete the electrochemical circuit.
15 Although it is not essential that a counter electrode be employed where a reference electrode is included in the circuit and the electrode is capable of performing the function of a counter electrode, it is preferred to have separate counter and reference electrodes because the
20 reference potential provided by the reference electrode is most stable when it is at equilibrium. If the reference electrode is required to act further as a counter electrode, the current flowing through the reference electrode may disturb this equilibrium.
25 Consequently, separate electrodes functioning as counter and reference electrodes are most preferred.

 In one embodiment, the "counter electrode" of the "sensor element" comprises a "bimodal electrode." The term "bimodal electrode" as used herein typically refers
30 to an electrode which is capable of functioning non-simultaneously as, for example, both the counter

electrode (of the "sensor element") and the iontophoretic electrode (of the "sampling means").

The terms "reactive surface," and "reactive face" are used interchangeably herein to mean the surface of the sensing electrode that: (1) is in contact with the surface of an electrolyte containing material (e.g. gel) which contains an analyte or through which an analyte, or a derivative thereof, flows from a source thereof; (2) is comprised of a catalytic material (e.g., carbon, platinum, palladium, rhodium, ruthenium, or nickel and/or oxides, dioxides and combinations or alloys thereof) or a material that provides sites for electrochemical reaction; (3) converts a chemical signal (e.g. hydrogen peroxide) into an electrical signal (e.g., an electrical current); and (4) defines the electrode surface area that, when composed of a reactive material, is sufficient to drive the electrochemical reaction at a rate sufficient to generate a detectable, reproducibly measurable, electrical signal that is correlatable with the amount of analyte present in the electrolyte.

The term "collection reservoir" is used to describe any suitable containment means for containing a sample extracted from a biological system. For example, the collection reservoir can be a receptacle containing a material which is ionically conductive (e.g., water with ions therein), or alternatively, it can be a material, such as, a sponge-like material or hydrophilic polymer, used to keep the water in place. Such collection reservoirs can be in the form of a hydrogel (for example, in the form of a disk or pad). Hydrogels are typically referred to as "collection inserts." Other

suitable collection reservoirs include, but are not limited to, tubes, vials, capillary collection devices, cannulas, and miniaturized etched, ablated or molded flow paths.

5 An "ionically conductive material" refers to any material that provides ionic conductivity, and through which electrochemically active species can diffuse. The ionically conductive material can be, for example, a solid, liquid, or semi-solid (e.g., in the form of a
10 gel) material that contains an electrolyte, which can be composed primarily of water and ions (e.g., sodium chloride), and generally comprises 50% or more water by weight. The material can be in the form of a gel, a sponge or pad (e.g., soaked with an electrolytic
15 solution), or any other material that can contain an electrolyte and allow passage therethrough of electrochemically active species, especially the analyte of interest.

 The term "physiological effect" encompasses effects
20 produced in the subject that achieve the intended purpose of a therapy. In preferred embodiments, a physiological effect means that the symptoms of the subject being treated are prevented or alleviated. For example, a physiological effect would be one that
25 results in the prolongation of survival in a patient.

 A "laminate" , as used herein, refers to structures comprised of at least two bonded layers. The layers may be bonded by welding or through the use of adhesives. Examples of welding include, but are not limited to, the
30 following: ultrasonic welding, heat bonding, and inductively coupled localized heating followed by localized flow. Examples of common adhesives include,

but are not limited to, pressure sensitive adhesives, thermoset adhesives, cyanoacrylate adhesives, epoxies, contact adhesives, and heat sensitive adhesives. An example of a laminate of the present invention is a mask
5 layer, collection inserts, and a retaining layer (e.g., Figure 3, 50) where at least the mask and retaining layer are bonded to each other.

A "collection assembly", as used herein, refers to structures comprised of several layers, where the
10 assembly includes at least one collection insert, for example a hydrogel. An example of a collection assembly of the present invention is a mask layer, collection inserts, and a retaining layer (e.g., Figure 3, 50) where the layers are held in appropriate, functional
15 relationship to each other but are not necessarily a laminate, i.e., the layers may not be bonded together. The layers may, for example, be held together by interlocking geometry or friction.

An "autosensor assembly", as used herein, refers to
20 structures generally comprising a mask layer, collection inserts, a retaining layer, an electrode assembly, and a support tray. The autosensor assembly may also include liners (e.g., the autosensor assembly shown in Figure 3) where the layers are held in appropriate, functional
25 relationship to each other.

The mask and retaining layers are preferably composed of materials that are substantially impermeable to the analyte (chemical signal) to be detected (e.g., glucose); however, the material can be permeable to
30 other substances. By "substantially impermeable" is meant that the material reduces or eliminates chemical signal transport (e.g., by diffusion). The material can

allow for a low level of chemical signal transport, with the proviso that chemical signal that passes through the material does not cause significant edge effects at the sensing electrode.

5 "Substantially planar" as used herein, includes a slightly curved surface that conforms, for example, to the curvature of the forearm or upper arm of a subject. A "substantially planar" surface is, for example, a surface having a shape to which skin can conform, i.e.,
10 creating contact between the skin and the surface. A further example includes shapes that have large length and width relative to their depth (e.g., 10:1 or greater) and permit the skin to conform to their surface topography.

15 By the term "printed" as used herein is meant a substantially uniform deposition of an electrode formulation onto one surface of a substrate (i.e., the base support). It will be appreciated by those skilled in the art that a variety of techniques may be used to
20 effect substantially uniform deposition of a material onto a substrate, e.g., Gravure-type printing, extrusion coating, screen coating, spraying, painting, or the like.

25 2. EXEMPLARY EMBODIMENTS OF SAMPLING SYSTEMS.

 The present invention relates to laminates, collection assemblies, and other components useful in a sampling device for transdermally extracting and
 measuring the concentration of a target analyte present
30 in a biological system. Such sampling devices are generally used for extracting small amounts of a target analyte from the biological system, and then sensing

and/or quantifying the concentration of the target analyte. Measurement and/or sampling with the sampling device can be carried out in a continual or continuous manner. Continual or continuous measurements allow for
5 closer monitoring of target analyte concentration fluctuations. In general, the sampling device comprises a biosensor with an electrochemical sensing element, and the sampling device is preferably used to perform
10 continual transdermal or transmucosal sampling of blood glucose.

The analyte can be any specific substance or component that one is desirous of detecting and/or measuring in a chemical, physical, enzymatic, or optical analysis. Such analytes include, but are not limited
15 to, amino acids, enzyme substrates or products indicating a disease state or condition, other markers of disease states or conditions, drugs of abuse, therapeutic and/or pharmacologic agents (e.g., theophylline, anti-HIV drugs, lithium, anti-epileptic
20 drugs, cyclosporin, chemotherapeutics), electrolytes, physiological analytes of interest (e.g., urate/uric acid, carbonate, calcium, potassium, sodium, chloride, bicarbonate (CO_2), glucose, urea (blood urea nitrogen), lactate/lactic acid, hydroxybutyrate, cholesterol,
25 triglycerides, creatine, creatinine, insulin, hematocrit, and hemoglobin), blood gases (carbon dioxide, oxygen, pH), lipids, heavy metals (e.g., lead, copper), and the like. In preferred embodiments, the analyte is a physiological analyte of interest, for
30 example glucose, or a chemical that has a physiological action, for example a drug or pharmacological agent.

In order to facilitate detection of the analyte, an enzyme can be disposed in the collection reservoir, or, if several collection reservoirs are used, the enzyme can be disposed in several or all of the reservoirs.

5 The selected enzyme is capable of catalyzing a reaction with the extracted analyte (in this case glucose) to the extent that a product of this reaction can be sensed, e.g., can be detected electrochemically from the generation of a current which current is detectable and
10 proportional to the concentration or amount of the analyte which is reacted. A suitable enzyme is glucose oxidase which oxidizes glucose to gluconic acid and hydrogen peroxide. The subsequent detection of hydrogen peroxide on an appropriate biosensor electrode generates
15 two electrons per hydrogen peroxide molecule which create a current which can be detected and related to the amount of glucose entering the device. Glucose oxidase (GOx) is readily available commercially and has well known catalytic characteristics. However, other
20 enzymes can also be used, so long as they specifically catalyze a reaction with an analyte or substance of interest to generate a detectable product in proportion to the amount of analyte so reacted.

In like manner, a number of other analyte-specific
25 enzyme systems can be used in the invention, which enzyme systems operate on much the same general techniques. For example, a biosensor electrode that detects hydrogen peroxide can be used to detect ethanol using an alcohol oxidase enzyme system, or similarly
30 uric acid with urate oxidase system, urea with a urease

system, cholesterol with a cholesterol oxidase system, and theophylline with a xanthine oxidase system.

In addition, the oxidase enzyme (used for hydrogen peroxide-based detection) can be replaced with another redox system, for example, the dehydrogenase-enzyme NAD-NADH, which offers a separate route to detecting additional analytes. Dehydrogenase-based sensors can use working electrodes made of gold or carbon (via mediated chemistry). Examples of analytes suitable for this type of monitoring include, but are not limited to, cholesterol, ethanol, hydroxybutyrate, phenylalanine, triglycerides, and urea. Further, the enzyme can be eliminated and detection can rely on direct electrochemical or potentiometric detection of an analyte. Such analytes include, without limitation, heavy metals (e.g., cobalt, iron, lead, nickel, zinc), oxygen, carbonate/carbon dioxide, chloride, fluoride, lithium, pH, potassium, sodium, and urea. Also, the sampling system described herein can be used for therapeutic drug monitoring, for example, monitoring anti-epileptic drugs (e.g., phenytion), chemotherapy (e.g., adriamycin), hyperactivity (e.g., ritalin), and anti-organ-rejection (e.g., cyclosporin).

More specifically, a non-invasive glucose monitoring (sampling) device is used to measure changes in glucose levels in an animal subject over a wide range of glucose concentrations. The sampling method is based on transdermal glucose extraction, and the sensing method is based on electrochemical detection technology. The device can be contacted with the biological system continuously, and automatically obtains glucose samples

in order to measure glucose concentration at various selected intervals.

Sampling is carried out continually by non-invasively extracting glucose through the skin of the patient. More particularly, an iontophoretic current is applied to a surface of the skin of a subject. When the current is applied, ions or charged molecules pull along other uncharged molecules or particles such as glucose which are drawn into a collection insert placed on the surface of the skin. The collection insert may comprise any ionically conductive material and is preferably in the form of a hydrogel which is comprised of a hydrophilic material, water and an electrolyte.

The collection insert may further contain an enzyme which catalyzes a reaction of glucose to form an easily detectable species. The enzyme is preferably glucose oxidase (GOx) which catalyzes the reaction between glucose and oxygen and results in the production of hydrogen peroxide. The hydrogen peroxide reacts at a catalytic surface of a biosensor electrode, resulting in the generation of electrons which create a detectable biosensor current (raw signal). Based on the amount of biosensor current created over a given period of time, a measurement is taken, which measurement is related to the amount of glucose drawn into the collection insert over a given period of time.

When the reaction is complete, the process can be repeated and a subsequent measurement obtained. More specifically, the iontophoretic current is again applied, glucose is drawn through the skin surface into the collection insert, and the reaction is catalyzed in order to create a biosensor current. These sampling

(extraction) and sensing operations can be integrated such that glucose is extracted into a hydrogel collection pad where it contacts the GOx enzyme. The GOx enzyme converts glucose and oxygen in the hydrogel to hydrogen peroxide which diffuses to the sensor and is catalyzed by the sensor to regenerate oxygen and form electrons. The electrons generate an electrical signal that can be measured, analyzed, and correlated to blood glucose.

10 In one embodiment of the present invention, the sampling system can have two collection reservoirs which contain, for example, an active collection reservoir, having the GOx enzyme, and a blank collection reservoir (without the GOx enzyme); or, in an alternative, two
15 active reservoirs, i.e., two reservoirs containing the GOx enzyme. In the case of an active collection reservoir and a blank collection reservoir signal can be adjusted by subtraction of the blank reservoir signal from the signal obtained from the active reservoir. In
20 the case of two active collection reservoirs the signals can be summed and averaged, or a total of the two signals can be used. This signal, for example the detected current, is then used alone or in combination with other factors (for example, glucose concentration
25 at a calibration point, skin temperature, conductivity, voltage, time since calibration of the system, etc.) to provide a glucose concentration value.

In particular embodiments, the detected current can be correlated with the subject's blood glucose
30 concentration (typically using statistical algorithms associated with a microprocessor) so that the system controller may display the subject's actual blood

glucose concentration as measured by the sampling system. For example, the system can be calibrated to the subject's actual blood glucose concentration by sampling the subject's blood during a standard glucose tolerance test, and analyzing the blood glucose using both a standard blood glucose monitor and the sampling system of the present invention. In addition or alternately, the sampling system can be calibrated at a calibration time point where the signal obtained from the sampling system at that time point is correlated to blood glucose concentration at that time point as determined by direct blood testing (for example, glucose concentration can be determined using a HemoCue® clinical analyzer (HemoCue AB, Sweden)). In this manner, measurements obtained by the sampling system can be correlated to actual values using known statistical techniques. Such statistical techniques can be formulated as algorithm(s) and incorporated in a microprocessor associated with the sampling system.

A generalized method for continual monitoring of a physiological analyte is disclosed in International Publication No. WO 97/24059, published 10 July 1997. As noted in that publication, the analyte is extracted into a reservoir containing a hydrogel which is preferably comprised of a hydrophilic material of the type described in International Publication No. WO 97/02811, published 30 January 1997. Suitable hydrogel materials include, but are not limited to, polyethylene oxide, polyacrylic acid, polyvinylalcohol and related hydrophilic polymeric materials combined with water to form an aqueous gel.

In the above non-invasive glucose monitoring device, a biosensor electrode is positioned against a surface of the hydrogel opposite the surface of the hydrogel which contacts the skin. The sensor electrode
5 acts as a detector which detects current generated by hydrogen peroxide in the redox reaction, or more specifically detects current which is generated by the electrons generated by the redox reaction catalyzed by the reactive surface of the electrode (International
10 Publication No. WO 96/00110, published 4 January 1996, and International Publication No. WO 97/10499, published 2 March 1997).

Referring now to Figures 1A and 1B, an exemplary iontophoretic collection reservoir and electrode
15 assembly for use in a transdermal sensing device is generally indicated at 2. The assembly comprises two iontophoretic collection reservoirs, 4 and 6, each generally comprising a conductive medium 8, and 10 (preferably cylindrical hydrogel pads), respectively
20 disposed therein. First (12) and second (14) ring-shaped iontophoretic electrodes are respectively contacted with conductive medium 8 and 10. The first iontophoretic electrode 12 surrounds three biosensor electrodes which are also contacted with the conductive
25 medium 8, a working electrode 16, a reference electrode 18, and a counter electrode 20. A guard ring 22 separates the biosensor electrodes from the iontophoretic electrode 12 to minimize noise from the iontophoretic circuit. Conductive contacts provide
30 communication between the electrodes and an associated power source and control means as described below. A similar biosensor electrode arrangement can be contacted

with the conductive medium 10, or the medium may not have a sensor means contacted therewith (e.g., in order to provide a blank).

Referring now to Figure 2, the iontophoretic collection reservoir and electrode assembly 2 of Figures 1A and 1B is shown in exploded view in combination with a suitable iontophoretic sampling device housing 32. The housing can be a plastic case or other suitable structure which preferably is configured to be worn on a subject's arm in a manner similar to a wrist watch. As can be seen, conductive media 8 and 10 (hydrogel pads) are separable from the assembly 2; however, when the assembly 2 and the housing 32 are combined to provide an operational iontophoretic sampling device 30, the media are in contact with the electrodes to provide a electrical contact therewith.

A power source (e.g., one or more rechargeable or nonrechargeable batteries) can be disposed within the housing 32 or within the straps 34 which hold the device in contact with a skin or mucosal surface of a subject. In use, an electric potential (either direct current or a more complex waveform) is applied between the two iontophoretic electrodes 12 and 14 such that current flows from the first iontophoretic electrode 12, through the first conductive medium 8 into the skin or mucosal surface, and then back out through the second conductive medium 10 to the second iontophoretic electrode 14. The current flow is sufficient to extract substances including an analyte of interest through the skin into one or both of collection reservoirs 4 and 6. The electric potential may be applied using any suitable

technique, for example, the applied current density may be in the range of about 0.01 to 0.5 mA/cm². In a preferred embodiment, the device is used for continual or continuous monitoring, and the polarity of

5 iontophoretic electrodes 12 and 14 is alternated at a rate of about one switch every 10 seconds to about one switch every hour so that each electrode is alternately a cathode or an anode. After a suitable iontophoretic extraction period, one or both of the sensor electrode

10 sets can be activated in order to detect extracted substances including the analyte of interest. Operation of the iontophoretic sampling device 30 is preferably controlled by a controller 36 (e.g., a microprocessor), which interfaces with the iontophoretic electrodes, the

15 sensor electrodes, the power supply, as well as optional temperature and/or conductance sensing elements, a display, and other electronics. For example, the controller 36 can include a programmable controlled circuit source/sink drive for driving the iontophoretic

20 electrodes. Power and reference voltage are provided to the sensor electrodes, and signal amplifiers can be used to process the signal from the working electrode or electrodes. In general, the controller discontinues the iontophoretic current drive during sensing periods.

25 In a further aspect of the above embodiments, the sensor element can also include a reference electrode, and a counter electrode. Further, a counter electrode of the sensor element and an iontophoretic electrode of the sampling system can be combined as a single bimodal

30 electrode where the electrode is not used simultaneously for both functions, i.e., where the counter and

iontophoretic functions are separately carried out at different times.

In one aspect, the sampling device can operate in an alternating polarity mode, for example, using first and second bimodal electrodes (Figure 7, 740 and 741) and two collection reservoirs (Figure 7, 747 and 748). Each bi-modal electrode (Figure 6, 630; Figure 7, 740 and 741) serves two functions depending on the phase of the operation: (1) an electro-osmotic electrode (or iontophoretic electrode) used to electrically draw analyte from a source into a collection reservoir comprising water and an electrolyte, and to the area of the electrode subassembly; and (2) as a counter electrode to the first sensing electrode at which the chemical compound is catalytically converted at the face of the sensing electrode to produce an electrical signal.

The reference (Figure 7, 744 and 745; Figure 6, 632) and sensing electrodes (Figure 7, 742 and 743; Figure 6, 631), as well as, the bimodal electrode (Figure 7, 740 and 741; Figure 6, 630) are connected to a standard potentiostat circuit during sensing. In general, practical limitations of the system require that the bimodal electrode will not act as both a counter and iontophoretic electrode simultaneously.

The general operation of an iontophoretic sampling system is the cyclical repetition of two phases: (1) a reverse-iontophoretic phase, followed by a (2) sensing phase. During the reverse iontophoretic phase, the first bimodal electrode (Figure 7, 740) acts as an iontophoretic cathode and the second bimodal electrode (Figure 7, 741) acts as an iontophoretic anode to

complete the circuit. Analyte is collected in the reservoirs, for example, a hydrogel (Figure 7, 747 and 748). At the end of the reverse iontophoretic phase, the iontophoretic current is turned off. During the sensing phase, in the case of glucose, a potential is applied between the reference electrode (Figure 7, 744) and the sensing electrode (Figure 7, 742). The chemical signal reacts catalytically on the catalytic face of the first sensing electrode (Figure 7, 742) producing an electrical current, while the first bi-modal electrode (Figure 7, 740) acts as a counter electrode to complete the electrical circuit.

The electrode described is particularly adapted for use in conjunction with a hydrogel collection reservoir system for monitoring glucose levels in a subject through the reaction of collected glucose with the enzyme glucose oxidase present in the hydrogel matrix.

The bi-modal electrode is preferably comprised of Ag/AgCl; other suitable substances are can be determined in view of the teachings of the present disclosure and the prior art. The electrochemical reaction which occurs at the surface of this electrode serves as a facile source or sink for electrical current. This property is especially important for the iontophoresis function of the electrode. Lacking this reaction, the iontophoresis current could cause the hydrolysis of water to occur at the iontophoresis electrodes causing pH changes and possible gas bubble formation. The pH changes to acidic or basic pH could cause skin irritation or burns. The ability of an Ag/AgCl electrode to easily act as a source of sink current is also an advantage for its counter electrode function.

For a three electrode electrochemical cell to function properly, the current generation capacity of the counter electrode should not limit the speed of the reaction at the sensing electrode. In the case of a large sensing electrode, the counter electrode should be able to source proportionately larger currents.

The design of the sampling system provides for a larger sensing electrode (see for example, Figure 6) than previously designed. Consequently, the size of the bimodal electrode should be sufficient so that when acting as a counter electrode with respect to the sensing electrode the counter electrode does not become limiting the rate of catalytic reaction at the sensing electrode catalytic surface.

Two methods exist to ensure that the counter electrode does not limit the current at the sensing electrode: (1) the bi-modal electrode is made much larger than the sensing electrode, or (2) a facile counter reaction is provided.

During the reverse iontophoretic phase, the power source provides a current flow to the first bi-modal electrode to facilitate the extraction of the chemical signal into the reservoir. During the sensing phase, the power source is used to provide voltage to the first sensing electrode to drive the conversion of chemical signal retained in reservoir to electrical signal at the catalytic face of the sensing electrode. The power source also maintains a fixed potential at the electrode where, for example, hydrogen peroxide is converted to molecular oxygen, hydrogen ions, and electrons, which is compared with the potential of the reference electrode during the sensing phase. While one sensing electrode

is operating in the sensing mode it is electrically connected to the adjacent bimodal electrode which acts as a counter electrode at which electrons generated at the sensing electrode are consumed.

5 The electrode sub-assembly can be operated by electrically connecting the bimodal electrodes such that each electrode is capable of functioning as both an iontophoretic electrode and counter electrode along with appropriate sensing electrode(s) and reference
10 electrode(s), to create standard potentiostat circuitry.

 A potentiostat is an electrical circuit used in electrochemical measurements in three electrode electrochemical cells. A potential is applied between the reference electrode and the sensing electrode. The
15 current generated at the sensing electrode flows through circuitry to the counter electrode (i.e., no current flows through the reference electrode to alter its equilibrium potential). Two independent potentiostat circuits can be used to operate the two biosensors. For
20 the purpose of the present sampling system, the electrical current measured at the sensing electrode subassembly is the current that is correlated with an amount of chemical signal.

 With regard to continual operation for extended
25 periods of time, Ag/AgCl electrodes are provided herein which are capable of repeatedly forming a reversible couple which operates without unwanted electrochemical side reactions (which could give rise to changes in pH, and liberation of hydrogen and oxygen due to water
30 hydrolysis). The Ag/AgCl electrodes of the present sampling system are thus formulated to withstand repeated cycles of current passage in the range of about

0.01 to 1.0 mA per cm² of electrode area. With regard to high electrochemical purity, the Ag/AgCl components are dispersed within a suitable polymer binder. One such example of a suitable binder is styrene acrylonitrile (SAN) to provide an electrode composition which is not susceptible to attack (e.g., plasticization) by components in the collection reservoir, e.g., the hydrogel composition. The electrode compositions are also formulated using analytical- or electronic-grade reagents and solvents, and the polymer binder composition is selected to be free of electrochemically active contaminants which could diffuse to the biosensor to produce a background current.

Since the Ag/AgCl iontophoretic electrodes must be capable of continual cycling over extended periods of time, the absolute amounts of Ag and AgCl available in the electrodes, and the overall Ag/AgCl availability ratio, can be adjusted to provide for the passage of high amounts of charge. Although not limiting in the sampling system described herein, the Ag/AgCl ratio can approach unity. In order to operate within the preferred system which uses a biosensor having a geometric area of 0.1 to 3 cm², the iontophoretic electrodes are configured to provide an approximate electrode area of 0.3 to 1.0 cm², preferably about 0.85 cm². These electrodes provide for reproducible, repeated cycles of charge passage at current densities ranging from about 0.01 to 1.0 mA/cm² of electrode area. More particularly, electrodes constructed according to the above formulation parameters, and having an approximate electrode area of 0.85 cm², are capable of a reproducible total charge passage (in both anodic and cathodic

directions) of 270 mC, at a current of about 0.3 mA (current density of 0.35 mA/cm²) for 48 cycles in a 24 hour period.

Once formulated, the Ag/AgCl electrode composition is affixed to a suitable rigid or flexible nonconductive surface (for example, polyester, polycarbonate, vinyl, acrylic, PETG (polyethylene terephthalate copolymer), PEN, and polyimide) as described above with respect to the biosensor electrode composition. A silver (Ag) underlayer is first applied to the surface in order to provide uniform conduction. The Ag/AgCl electrode composition is then applied over the Ag underlayer in any suitable pattern or geometry using various thin film techniques, such as sputtering, evaporation, vapor phase deposition, or the like, or using various thick film techniques, such as film laminating, electroplating, or the like. Alternatively, the Ag/AgCl composition can be applied using screen printing, pad printing, inkjet methods, transfer roll printing, or similar techniques. Preferably, both the Ag underlayer and the Ag/AgCl electrode are applied using a low temperature screen print onto a polymeric substrate, for example, polyester. This low temperature screen print can be carried out at about 125 to 160°C, and the screening can be carried out using a suitable mesh, ranging from about 100-400 mesh.

In one embodiment, the electrode assemblies can include bimodal electrodes as shown in Figure 6 and described above.

The components described herein are intended for use in a automatic sampling device which is configured to be worn like an ordinary wristwatch. As described in

International Publication No. WO 96/00110, published 4 January 1996, the wristwatch housing typically contains conductive leads which communicate with the iontophoretic electrodes and the biosensor electrodes to control cycling and provide power to the iontophoretic electrodes, and to detect electrochemical signals produced at the biosensor electrode surfaces. The wristwatch housing can further include suitable electronics (e.g., microprocessor, memory, display and other circuit components) and power sources for operating the automatic sampling system.

Further, the sampling system can be pre-programmed to begin execution of its signal measurements (or other functions) at a designated time. One application of this feature is to have the sampling system in contact with a subject and to program the sampling system to begin sequence execution during the night so that it is available for calibration immediately upon waking. One advantage of this feature is that it removes any need to wait for the sampling system to warm-up before calibrating it.

3. LAMINATES, COLLECTION ASSEMBLIES, AND AUTOSENSOR ASSEMBLIES

The present invention relates to laminates, collection assemblies, and autosensors for use in a sampling device. More particularly, the present laminates, collection assemblies, and autosensors are used in a transdermal sampling device that is placed in operative contact with a skin or mucosal surface of the biological system to obtain a chemical signal associated with an analyte of interest. The sampling device

transdermally extracts the analyte from the biological system using, for example, an iontophoretic sampling technique. The transdermal sampling device can be maintained in operative contact with the skin or mucosal surface of the biological system to provide such
5 continual or continuous analyte measurement.

In one aspect, the invention relates to a collection assembly for use in a iontophoretic sampling device useful to monitor a selected analyte, or
10 derivatives thereof, present in a biological system. The collection assembly can include:

a) a mask layer comprised of a substantially planar material that is substantially impermeable to the selected analyte or derivatives thereof, where the mask
15 layer has inner and outer faces and the outer face provides contact with the biological system;

b) a collection insert layer comprised of an ionically conductive material having first and second surfaces, and

20 c) the mask and collection insert layers are configured such that (i) at least a portion of the collection insert is exposed to provide contact with the biological system, and (ii) flow of the analyte through the first surface of the collection insert layer from
25 the biological system is prevented by the mask layer for any portion of the first surface of the collection insert layer that is in contact with the inner face of the mask layer. Such collection assemblies can be included in autosensor assemblies typically including
30 (a) the collection assembly, (b) an electrode assembly

having an inner face comprising an electrode and an outer face, where the inner face of the electrode assembly and the collection assembly are aligned to define a plurality of flow paths through said collection assembly, and (c) a support tray that contacts the outer face of the electrode assembly. One example of this type of collection assembly and autosensor assembly is described in Example 2.

In a further aspect, the invention includes a collection assembly having a mask layer, a collection insert layer comprised of an ionically conductive material, wherein the layers are axially aligned to provide a flow path through the collection assembly. Typically, the mask layer is comprised of a material that is substantially impermeable to the chemical signal associated with the analyte of interest. Exemplary embodiments of such collection assemblies are described in Examples 1 and 2. Example 1 describes use of a retaining layer as well.

In one embodiment, the mask layer and retaining layer each define at least one opening and at least a portion of a collection insert is exposed by each opening to provide a flow path through the collection assembly. Further, the collection insert may be contained by a corral or gasket that contains, seals, or retains the collection insert at a desired location. When a gasket is used the entire surface of the collection insert may be exposed, for example, by the mask layer. In this case the mask layer contacts the edges of the gasket.

In another embodiment, the mask layer and retaining layer each define two openings and at least a portion of a collection insert is exposed by each opening to provide two flow paths through the collection assembly. As stated above, the collection inserts may each be contained by a corral or gasket.

The mask layer may be coated with an adhesive on either of its faces or on both of its faces. Further, a liner may be adhered to one of the faces of the mask layer, typically the outer face. Similarly for the retaining layer. In one embodiment, (i) the outer face of the mask layer has an adhesive coating and a liner attached, (ii) the inner face of the mask layer contacts the collection inserts and adheres to the inner face of the retaining layer, and (iii) the outer face of the retaining layer is adhered to a second liner (e.g., a plow-fold liner).

The collection assemblies may be prepared as laminates. Further, other components, such as support trays and electrodes or electrode assemblies can be combined with the collection assemblies or laminates to form autosensor assemblies.

Further, the collection assemblies, laminates, and autosensors of the invention may be provided in sealed packages. Such sealed packets may further comprise a source of hydration (e.g., a hydrating insert) which ensures that the collection inserts will not dehydrate prior to use.

The collection assemblies, laminates, and autosensors of the present invention are particularly well suited for use as consumable components in the

iontophoretic sampling device of Figure 2. Referring now to Figure 3, one embodiment of a collection assembly for use in such a sampling device is generally indicated at 50. The assembly is aligned with an electrode assembly 60 which includes both iontophoretic 59 and sensing electrodes 61 as described above. A tray 70 is adapted to hold the electrode and collection assemblies in operative alignment, and provides electrical connection between the electrode assembly and control components provided by an associated housing element (e.g., housing 32 of Figure 2). If desired, the tray 70 can be comprised of a substantially rigid substrate and have features or structures which cooperate and/or help align the various assemblies in the sampling device.

For example, the tray can have one or more wells or recesses, and/or one or more lips, rims, or other structures which depend from the substrate, each of which features or structures facilitate register between the electrode assembly, the collection assembly and the associated components of the sampling device. The tray can be composed of any suitable material, desirable characteristics of which can include the following: (i) high heat distortion temperature (to allow hot melt bonding of the electrode assembly to the tray, if necessary or desired); (ii) optimum rigidity, to allow for ease of handling and insertion into the housing of the monitoring device; (iii) low moisture uptake, to insure that proper hydration of the ionically conductive medium (e.g., hydrogel collection inserts) is maintained when the medium is stored in proximity to the tray; and, (iv) moldable by conventional processing techniques, for example, injection molding.

Materials for use in manufacturing the tray include, but are not limited to, the following: PETG (polyethylene terephthalate copolymer); ABS (acrylonitrile-butadiene-styrene co-polymer); SAN (styrene-acrylonitrile copolymer); SMA (styrene-maleic anhydride copolymer); HIPS (high impact polystyrene); polyethylene terephthalate (PET); polystyrene (PS); polypropylene (PP); and blends thereof. In a preferred embodiment the tray is formed from high impact polystyrene.

The electrode assembly is typically fixed to the tray to, for example, facilitate register between the electrode assembly and the associated components of the housing of the sampling device. The electrode assembly may be manufactured as part of the tray, or, the electrode assembly may be attached to the tray by, for example, (i) using connecting means which allow the electrode assembly to engage the tray (e.g., holes in the electrode assembly with corresponding pegs on the tray); or (ii) use of an adhesive. Exemplary adhesives include, but are not limited to, the following: acrylate, cyanoacrylate, styrene-butadiene, co-polymer based adhesives, and silicone. In a preferred embodiment the tray is attached to the electrode assembly as in (i) above with the pegs deformed, thus locking the components together.

The collection assembly 50 includes one or more collection inserts 52 that are comprised of an ionically conductive material. Each collection insert has first and second opposing surfaces, 54 and 56, respectively. The collection insert is preferably comprised of a substantially planar hydrogel disk. The first opposing

surface 54 of the insert is intended for contact with a target surface (skin or mucosa), and the second opposing surface 56 is intended for contact with the electrode assembly 60, thereby establishing a flow path between

5 the target surface and the iontophoretic and sensing electrodes. A mask layer 58 is positioned over the first surface 54 of the collection insert. The mask layer is used to inhibit contact between the sensing electrode(s) of the electrode assembly and chemical

10 signal that may be transported in a radial direction from the target surface. The mask layer 58 comprises at least one opening 62 which is sized to allow a detectable amount of chemical signal to reach the sensing electrode, while reducing or preventing entry of

15 chemical signal into the flow path thorough the insert that has a potential to be transported (e.g., by diffusion) in a radial direction toward an edge of the sensing electrode. As explained in commonly owned U.S. Patent No. 5,735,273, this type of mask layer serves to

20 substantially eliminate "edge effect" flow, i.e., the mask prevents chemical signal from contacting the electrode unless the signal flows substantially perpendicular to the surface of the sensing electrode. Accordingly, the opening 62 in the mask layer is sized

25 to expose at least a portion of the first surface 54 of the collection insert. In the particular embodiment depicted in Figure 3, a border region 66 of the mask layer generally extends beyond the first surface of the collection insert to provide an overhang.

30 A retaining layer 68 is positioned in facing relation with the second surface 56 of the collection

insert 52. The retaining layer has at least one opening 72 which exposes at least a portion of the second surface 56 of the collection insert. Again, in the particular embodiment of Figure 3, a border region 74 of the retaining layer 68 extends beyond the second surface 56 in order to provide an overhang. The overhangs provided by the mask and retaining layers serve as a point of attachment between the two layers. When these layers are attached to each other at their overhanging portions, a laminate is formed wherein the collection insert is sandwiched between the two layers to provide a three-layer structure. Although the overhangs provided by border regions 66 and 74 are depicted in Figure 3 as extending along an edge of the mask and retaining layers, the overhangs can, of course, be formed from one or more corresponding tab overhangs (positioned anywhere on the subject layers), one or more corresponding edges (opposite and/or adjacent edges), or can be formed from a continuous overhang which encompasses the collection insert (e.g., an overhang which circumscribes an oval- or circular-shaped insert, or an overhang which surrounds all sides of a square-, rectangular-, rhomboid-, or triangular-shaped insert).

The one or more openings 62 in the mask layer, and the one or more openings 72 in the retaining layer can have any suitable geometry which is generally dictated by the shape of the collection insert 52 and/or the shape of the iontophoretic and sensing electrodes 59 and 61 used in the electrode assembly 60. In the embodiment depicted in Figure 3, wherein the electrodes are arranged in a circular configuration and the collection

insert is a circular disk, openings 62 and 72 preferably have a round, oval, ellipsoid, or "D"-shape which serves to collimate the flow (i.e., reduce or eliminate the edge effect flow) of chemical signal as it passes
5 through the collection assembly toward the electrode assembly 60.

The openings 62 and 72 in the mask and retaining layers can be sized the same or differently, wherein the particular sizes of the openings are generally set by
10 the overall surface area of the sensing electrode 61 that the collection assembly must operate with in the sensing device. Although the collection assemblies of the present invention can be provided in any size suitable for a targeted skin or mucosal surface, an
15 assembly that is used with a sampling device that contacts a subject's wrist will generally have a surface area on each face in the range of about 0.5 cm² to 15 cm². The openings 62 and 72 generally expose about 50% of the area of the sensing electrode, within a
20 manufacturing tolerance of about $\pm 20\%$. In general, the openings constitute an area that is in the range of 1% to 90% of the surface area encompassed by the mask or retaining layer plus the opening(s). The openings are, however, sized smaller than the overall surface of the
25 collection insert in at least one dimension.

The size or geometric surface area of the sensing electrode 61, the thickness of the collection insert 52, the sizes of the openings 62 and 72 in the mask and retaining layers, and the size of the overhangs provided
30 by border regions 66 and 74 of the mask and retaining layers are all interrelated to each other. For example, when the thickness of the collection insert is

increased, the size of the opening is decreased to obtain the same degree of reduction of edge effect flow (radial transport) of the transported chemical signal. Any decrease in the size of the openings in the mask and retaining layers increases the ability to block such unwanted flow. However, it is also desirable to maximize the size of the openings in order to maximize the amount of chemical signal which contacts the reactive surface of the sensing electrode 61.

10 The physical characteristics of the mask and retaining layers are selected so as to optimize the operational performance of the collection assembly. More particularly, since the assembly is intended to be contacted with a target surface for an extended period of time, the layers preferably have sufficient mechanical integrity so as to provide for such extended use. Furthermore, the layers should have sufficient flex and stretchability so as to resist tearing or rupture due to ordinary motion in the target surface, for example, movement of a subjects arm when the sampling device is contacted with a forearm or wrist. The layers can also have, for example, rounded corners which tolerate a greater degree of twist and flex in a target area (without breaking contact) than layers which have sharp, angular corners. The layers also provide for some degree of sealing between the target surface and the collection assembly 50, and between the collection assembly and the electrode assembly 60, and can provide for electrical, chemical, and/or electrochemical isolation between multiple collection inserts in the collection assembly and their corresponding electrodes in the electrode assembly.

Other physical characteristics include the degree of occlusivity provided by the mask layer, adhesion to the target surface and/or electrode assembly, and mechanical containment of the associated collection insert(s). In one embodiment, the collection assembly includes two collection inserts (as depicted in Figure 3), and the mask and retaining layers have corresponding central regions, 76 and 78, respectively, which are disposed between corresponding openings in the layers and provide for a further point of attachment between the two layers. As will be appreciated by the skilled artisan upon reading the present specification, this further point of attachment provides for chemical and electrical isolation between the two collection inserts.

The mask and retaining layers are preferably composed of materials that are substantially impermeable to the analyte (chemical signal) to be detected (e.g., glucose); however, the material can be permeable to other substances. By "substantially impermeable" is meant that the material reduces or eliminates chemical signal transport (e.g., by diffusion). The material can allow for a low level of chemical signal transport, with the proviso that chemical signal that passes through the material does not cause significant edge effects at the sensing electrode used in conjunction with the mask and retaining layers. Examples of materials that can be used to form the layers include, without limitation, the following: polymeric materials -- such as, polyethylene (PE) {including, high density polyethylene (HDPE), low density polyethylene (LDPE), and very low density polyethylene (VLDPE)}, polyethylene copolymers, thermoplastic elastomers, silicon elastomers,

polyurethane (PU), polypropylene (PP), (PET), nylon, flexible polyvinylchloride (PVC), and the like; natural rubber or synthetic rubber, such as latex; and combinations of the foregoing materials. Of these materials, exemplary flexible materials include, but are not limited to, the following: HDPE, LDPE, nylon, PET, PP, and flexible PVC. Stretchable materials include, but are not limited to, VLDPE, PU, silicone elastomers, and rubbers (e.g., natural rubbers, synthetic rubbers, and latex). In addition, adhesive materials, for example, acrylate, styrene butadiene rubber (SBR) based adhesives, styrene-ethylene-butylene rubber (SER) based adhesives, and similar pressure sensitive adhesives, can be used to form layers as well.

Each layer can be composed of a single material, or can be composed of two or more materials (e.g., multiple layers of the same or different materials) to form a chemical signal-impermeable composition.

Use of a mask to reduce or eliminate chemical signal which can radially transport toward a working electrode was described in co-owned U.S. Patent Nos. 5,735,273 and 5,827,183.

Methods for making the mask and retaining layers include, without limitation, extrusion processes, flow and form molding techniques, die cutting, and stamping techniques, which are all practiced according to methods well known in the art. Most preferably, the layers are manufactured in a manner that is the most economical without compromising performance (e.g., impermeability to a chemical signal, the ability to manipulate the layers by hand without breaking or otherwise compromising operability, and the like). The layers may

further have an adhesive coating (e.g., a pressure sensitive adhesive) on one or both surfaces. Exemplary adhesives include, but are not limited to, the following: starch, acrylate, styrene butadiene rubber-based, silicone, and the like. Adhesives that may come in contact with skin have a toxological profile compatible with skin-contact. In an exemplary embodiment, SBR-adhesive RP100 (John Deal Corporation, Mount Juliet, TN) can be used on both sides of a 0.001 inch thick PET film (Melinex #329, DuPont) retaining layer to adhere to the mask and the other side to the sensor. Another exemplary embodiment uses acrylate #87-2196 (National Starch and Chemical Corporation, Bridgewater, NJ) on the skin side of a 0.002 inch thick polyurethane (e.g., Dow Pellethane; Dow Chemical Corp., Midland, MI) mask to adhere the mask to the skin. Further, the mask and retaining layers may be coated with a material which absorbs one or more compounds or ions that may be extracted into the collection insert during sampling.

Since the collection assemblies of the present invention are intended for use as consumable (replaceable) components for a sampling device, the various constituents of the assemblies are preferably manufactured and then pre-assembled in an easy-to-use laminate structure that can be inserted and then removed from the sampling device housing by the consumer. In this regard, after the mask layer 58, retaining layer 68, and collection insert(s) 56 are produced, they are aligned as shown in Figure 3, and the overhangs provided by borders 66 and 74 are attached to each other to provide a three-layer laminate which sandwiches the

collection insert in between the mask and retaining layers as described above. The laminate, or a plurality of such laminates can be provided in a sealed package in order to maintain the cleanliness of the collection assembly (e.g., avoid chemical contamination from manufacturer and/or consumer handling) prior to use, and further to avoid dehydration of the collection inserts prior to use.

If desired, the package can include a source of hydration (e.g., a hydrating insert formed from a water-soaked pad, non-woven material, or gel which ensures that the collection inserts will not dehydrate prior to use. The hydrating insert may include other components as well, such as, buffers and antimicrobial compounds. The source of hydration is disposed of after the laminate has been removed from the package, and thus does not typically form a component of the sampling device.

The pre-assembled collection assembly laminates can include one or more optional liners which facilitate handling of the assembly. For example, a removable liner 80 can be applied over the mask layer 58, particularly when the mask layer is coated with an adhesive. An additional removable liner 90 can be applied over the retaining layer 68. The removable liners 80 and 90 are intended to remain in place until just prior to use of the assembly, and are thus manufactured from any suitable material which will not be too difficult to remove, but which will remain in place during packaging, shipment and storage to provide added protection to the assembly. If the mask and/or retaining layers are coated with (or actually formed

from) an adhesive, the removable liners can preferably be comprised of a polypropylene or treated polyester material which does not adhere well to commonly used contact adhesives. Other suitable materials include, without limitation, water and/or solvent impermeable polymers (including, but not limited to PET, PP, PE, and the like) and treated metal foils.

The removable liners 80 and 90 are generally shaped to cover the outer surfaces of the mask and retaining layers. The liners can further include grasping means, such as the tab 82 depicted in Figure 3, and intuitive indicia (such as numbering) which indicates the order in which the liners are intended to be removed during assembly of the sampling device. If desired, the liners can be shaped in a folded "V" (i.e., a "plow-fold" liner, see, e.g., liner 90 of Figure 3) or "Z" shape which provides a grasping means for the user, as well as providing for a controlled release motion in the liner. Alternatively, the liners can have an internal cut (e.g., a spiral cut extending from one edge of the liner and ending in the surface of the liner) or a scoring pattern which facilitates removal of the liner. Particularly, the liner material, shape, and related cuts or patterns or weakness are selected to ensure that removal of the liners does not delaminate the collection assembly, or otherwise disrupt the alignment between the various components of the collection assembly (i.e., the alignment between the mask layer, retaining layer, and the collection insert).

Production of one embodiment of the above-described collection assembly and autosensor assembly is presented in Example 1.

Referring now to Figure 4, a related embodiment of a collection assembly produced according to the present invention is generally indicated at 100. The assembly 100 is aligned with an electrode assembly 110 which
5 includes iontophoretic 109 and sensing electrodes 111 as described above, and is adapted to be held by a tray 120 as also described above. The collection assembly 100 includes one or more collection inserts 102 that are comprised of an ionically conductive material, and each
10 collection insert has first and second opposing surfaces, 104 and 106, respectively.

The first opposing surface 104 of the collection insert 102 is intended for contact with a target surface (skin or mucosa), and the second opposing surface 106 is
15 intended for contact with the electrode assembly 110, thereby establishing a flow path between the target surface and the iontophoretic and sensing electrodes. As above, a mask layer 108 is positioned over the first surface 104 of the collection insert, and includes one
20 or more openings 112 which provide for a collimated flow path between the target surface and the electrode assembly as also described above. The opening 112 in the mask layer 108 is sized smaller in at least one dimension relative to the surface area of the collection
25 insert 102.

A top surface 124 of a second layer 118 is positioned in facing relation with the bottom surface 114 of mask layer 108. The second layer comprises a gasket which has at least one opening 122. A two-layer
30 laminate is formed when the mask and second layers are attached at their respective facing surfaces. The

second layer also includes the collection insert 102 which is disposed within, and substantially fills the opening 122.

5 The physical and material properties of the mask layer are substantially identical to those of the mask layer described hereinabove, and the size and shape of the one or more openings are also determined using the above selection criteria. Furthermore, techniques for manufacture and manipulation of the mask layer 108 are
10 substantially identical to those techniques described above. However, unlike the above-described retaining layer, the gasket in the second layer 118 of the present embodiment is intended to serve as a corral for the collection insert. More particularly, the gasket
15 maintains the collection insert in a particular orientation such that, when the collection assembly is combined (contacted) with the electrode assembly, the collection insert is properly aligned with the iontophoretic and sensing electrodes. The gasket
20 material further provides for electrical and/or chemical isolation between multiple collection inserts, and provides structure to the collection assembly.

The second layer gasket can be formed from any suitable material such as those materials used in the
25 mask and retaining layers of the present invention. The gasket material could be a foam material that is sized to fit within the dimensions of the tray 120. Exemplary gasket materials include, without limitation, PE, PP, PET, nylon, and foamed PE. The gasket material can
30 further have an adhesive coating or layer which contacts the electrode assembly and provides for the facile

alignment between the electrode and collection assemblies.

Optional release liners 130 and/or 140 (a plow-fold liner) can also be respectively applied against the mask
5 layer 108 and second layer 118 to facilitate handling of the collection assemblies as described above. Furthermore, pre-assembled collection assembly laminates are preferably packaged, either individually or in groups, as also previously described.

10 Referring now to Figure 5, a still further related embodiment of a sampling system collection assembly is generally indicated at 150. The collection assembly 150 is aligned with an electrode assembly 160 which includes iontophoretic 159 and sensing electrodes 161 as
15 described above, and is adapted to be held by a tray 170. The collection assembly 150 includes one or more collection inserts 152 that are comprised of an ionically conductive material, and each collection insert has first and second opposing surfaces, 154 and
20 156, respectively.

The first opposing surface 154 of the collection insert 152 is intended for contact with a target surface (skin or mucosa), and the second opposing surface 156 is intended for contact with the electrode assembly 120,
25 thereby establishing a flow path between the target surface and the iontophoretic and sensing electrodes. As above, a mask layer 158 is positioned over the first surface 154 of the collection insert, and includes one or more openings 162 which provide for a collimated flow
30 path between the target surface and the electrode assembly as also described above. The opening 162 in

the mask layer 158 is sized smaller in at least one dimension relative to the surface area of the collection insert 152.

5 A top surface 174 of a second layer 168 is positioned in facing relation with the bottom surface 164 of the mask layer 158. The second layer comprises a gasket which has at least one opening 172. The second layer also includes the collection insert 152 which is disposed within, and substantially fills the opening
10 172.

The collection assembly 150 further includes a retaining layer 178, having a top surface 180 that is positioned in facing relation with the bottom surface 176 of the second layer 168. The retaining layer has at
15 least one opening 182 which exposes at least a portion of the second surface 156 of the collection insert 152. When the corresponding surfaces of the mask layer and second layer are attached to each other, and the corresponding surfaces of the second layer and the
20 retaining layer are attached to each other, a laminate is formed wherein both the second layer and the collection insert are sandwiched between the mask and retaining layers to provide a three-layer structure.

The physical and material properties of the mask
25 and retaining layers are substantially identical to those of the mask and retaining layers described hereinabove, and the size and shape of the one or more openings are also determined using the above selection criteria. Furthermore, techniques for manufacture and
30 manipulation of the mask and retaining layers 158 and 178 are substantially identical to those techniques

described above. Furthermore, the physical and material properties of the second layer gasket are substantially identical to those described above.

Optional release liners can also be applied against the mask layer 158 and retaining layer 178 to facilitate handling of the collection assemblies as described above. Furthermore, pre-assembled collection assembly laminates are preferably packaged, either individually or in groups, as also previously described.

EXPERIMENTAL

It is to be understood that while the invention has been described in conjunction with the preferred specific embodiments thereof, that the description above as well as the examples which follow are intended to illustrate and not limit the scope of the invention. Other aspects, advantages and modifications within the scope of the invention will be apparent to those skilled in the art to which the invention pertains.

In the following examples, efforts have been made to ensure accuracy with respect to numbers used (e.g., amounts, temperature, etc.) but some experimental error and deviation should be accounted for. Unless indicated otherwise, temperature is in degrees C and pressure is at or near atmospheric.

Example 1

An Exemplary Autosensor Assembly

A tray was produced using a high impact polystyrene (e.g., Chevron Valtra HG200N02; Chevron Chemical Corp., Houston, TX) in a plastic injection molding process. General shape and dimensions of the tray are indicated

in Figure 8A (the tray was 0.110 inches thick, with raised areas indicated in the figure). Dimensions in Figures 8A to 8H are all given in inches.

An electrode assembly was produced using thick film
5 ink formulations in a screen printing process. Each ink formulation comprised: a) an electrically conductive particulate, b) an electrochemically active particulate, c) a polymeric binder, and d) a volatile organic solvent to create a liquid slurry. During screen printing, the
10 inks were patterned onto the polyethylene terephthalate (PET; e.g., Melinex ST507, Dupont deNemours, Wilmington, DE) substrate and dried in place by passing through convection ovens. General shape and dimensions of the electrode assembly are indicated in Figures 8B (the
15 sensor is shown as lying flat for clarity; the material was polymer thick film inks on a 0.005 inch thick PET substrate).

The tray and electrode assembly were aligned using precisely punched holes in the sensor substrate that
20 engage with molded-in pins in the tray. The pins were plastically deformed (staked) with a blunt metal punch to fix the sensor substrate to the tray.

A tri-layer laminate was produced as follows (Figures 8C, 8D, and 8E). A mask layer, having the
25 general shape and dimensions shown in Figure 8C, was produced from a 0.002 inch thick polyurethane film (e.g., Dow Pellethane #2363-80AE; Dow Chemical Corp., Midland, MI) coated on one side with an 0.001 inch layer of acrylic pressure sensitive adhesive (e.g., Duro-tak,
30 #87-2196; National Starch and Chemical Corporation, Bridgewater, NJ). A rotary die cutting press was used to create the circular openings, to create the outline

border/perimeter geometry, and to laminate the mask material to the patient liner roll stock (0.003 inch thick PET coated on one side with silicone release; e.g., Fox River #1730 (Fox River Associates, Geneva, IL)).

The collection inserts were two essentially circular hydrogel disks, illustrated in Figure 8D, made from a water solution of polyethylene oxide, phosphate buffer, and glucose oxidase, impregnated in a 0.004 inch thick nonwoven PET (e.g., Remy™ #2250). This composite began as roll stock from which circular discs were cut and placed into contact with the mask material using a male-female punch set.

A retaining layer, having the general shape and dimensions shown in Figure 8E, was produced from 0.001 inch thick PET film (e.g., DuPont Melinex #329; Dupont deNemours, Wilmington, DE) coated on both sides with a styrene-butadiene-based pressure sensitive adhesive (e.g., RP100; John Deal Corporation, Mount Juliet, TN).

A rotary die cutting press was used to create the circular openings and outline border/perimeter geometry. A laminating press was used to place the retaining layer in contact with the collection insert and mask.

The openings in the mask layer were sized to expose a portion of the surface of each collection insert. A border region of the mask layer extended beyond the first surfaces of the collection insert to provide an overhang. The retaining layer was positioned in facing relation with the second surfaces of the collection insert. The retaining layer had two openings which exposes portions of the second surfaces of the collection insert. A border region of the retaining

layer extended beyond the second surfaces of the collection insert in order to provide an overhang. The overhanging portions of the mask and retaining layer served as points of attachment where the retaining layer
5 adhesive bound to the non-adhesive surface of the mask and thus prevented movement. This attaching of the layers to each other at their overhanging portions created a laminate where the collection insert was sandwiched between the two layers to provide a three-
10 layer structure.

The mask layer perimeter extended beyond the retaining layer perimeter, thus creating a third overhang. This overhang allows the mask layer to conform to the contours of the biological system to
15 which it is contacted (for example, a human forearm) and to be unencumbered by the rigidity of other parts of the autosensor assembly (for example, the tray and electrodes). Superior adhesion and reduced irritation were achieved by employing such an overhang.

20 The outline geometry of the patient liner, shown in Figure 8F, was produced during a blanking operation that used steel rule dies to cut the patient liner roll stock.

A second liner, i.e., a plow-fold liner, for
25 contacting the retaining layer and collection inserts was produced from 0.0016 inch thick biaxially oriented polypropylene film coated on one side with silicone release (e.g., Fox River, #1803; Fox River Associates). The treated side of the plowfold liner faced the
30 collection inserts, retaining layer, and sensor. This film was folded and perforated to length on a rotary press. The folded film was pulled apart at its

perforations to create single liners and was laminated to the outer adhesive surface of the retaining layer (dimensions shown in Figure 8G).

5 The plowfold liner, as described, left a portion of the retaining layer adhesive exposed. This adhesive was pressed into contact with the electrode-to-tray assembly during the plowfold lamination process, thus adhering the tri-layer laminate with liners to the electrode-to-tray assembly. A custom-designed assembly machine
10 performed the lamination using fixtures to precisely align the components relative to each other. Each component part nested precisely within its respective fixture to provide the necessary alignment. Vacuum was used to keep the parts from falling out of their
15 fixtures during assembly.

The entire assembly described above (Figure 8H), including, the tray, electrode assembly, tri-layer laminate, and liners, comprises an exemplary autosensor assembly of the present invention.

20 This particular embodiment of the autosensor assembly is also graphically represented in Figure 3 and Figures 8A-8H and is intended for use in the Glucowatch® biographer (Cygnus, Inc., Redwood City, CA), an iontophoretic sampling system for glucose concentration
25 monitoring of a subject.

Alternative materials for the components described above include, but are not limited to, the following:

(i) Alternate Mask, Retaining Layer, Patient
Liner, and Plowfold Liner Materials: high density
30 polyethylene (HDPE), low density polyethylene (LDPE), very low density polyethylene (VLDPE), polyethylene copolymers, thermoplastic elastomers, silicon

elastomers, polyurethane (PU), polypropylene (PP), (PET), nylon, flexible polyvinylchloride (PVC), natural rubber, synthetic rubber, and suitable combinations of the foregoing materials.

5 (ii) Base films: polyurethane, polyethylene (high density, medium density, low density, very low density, linear low density, very low density linear), polyethylene terephthalate (PET, polyester), vinyl, polystyrene, polycarbonate, diacetate, paper products,
10 blends of these materials, foam films made from any of the same materials;

(iii) Adhesives: acrylic based pressure-sensitive, rubber based pressure-sensitive (for example, styrene butadiene rubber (SBR) based adhesives, styrene-
15 ethylene-butylene rubber (SEBR) based adhesives), cyanoacrylates, epoxies, acrylate, and other pressure sensitive adhesives.

(iv) Release coatings: silicone, fluorinated polymers, chlorinated polymers; and

20 (v) Alternate tray materials: polycarbonate; PETG (polyethylene terephthalate copolymer); ABS (acrylonitrile-butadiene-styrene co-polymer); SAN (styrene-acrylonitrile copolymer); SMA (styrene-maleic anhydride copolymer); HIPS (high impact polystyrene);
25 polyethylene terephthalate (PET); polystyrene (PS); polypropylene (PP); and blends thereof.

Example 2

Another Embodiment of the Autosensor Assembly

30 A tray is produced using a high impact polystyrene (e.g., Chevron HG200N02) in a plastic injection molding process. General shape and dimensions of the tray are

indicated in Figure 9A (the tray was 0.110 inches thick, with raised areas indicated in the figure). Dimensions in all Figures 9A to 9G are given in inches.

Sensor-to-tray is assembled as described in Example 1. All other components are made by cutting from sheet stock using still rule dies, and hand assembled using visual alignment.

An electrode assembly is produced using thick film ink formulations in a screen printing process. Each ink formulation comprises: a) an electrically conductive particulate b) an electrochemically active particulate, c) a polymeric binder, and d) a volatile organic solvent to create a liquid slurry. During screen printing, the inks are patterned onto the polyethylene terephthalate (PET) substrate and dried in place by passing through convection ovens. General shape and dimensions of the electrode assembly are indicated in Figure 9B (the sensor is shown as lying flat for clarity; the material is polymer thick film inks on a 0.005 inch thick PET substrate).

The tray and electrode assembly are aligned using precisely punched holes in the sensor substrate that engage with molded-in pins in the tray. The pins are plastically deformed (staked) with a blunt metal punch to fix the sensor substrate to the tray.

A mask layer having the general shape and dimensions shown in Figure 9C is produced (steel-rule die cut from sheet stock) from 0.0015 inch Deerfield natural HDPE coated on two side with 0.001 inch adhesive (e.g., Duro-tak™, #87-2196; National Starch and Chemical Corporation, Bridgewater, NJ).

The collection inserts are two essentially circular hydrogel disks, illustrated in Figure 9D, made from a water solution of polyethylene oxide, phosphate buffer, and glucose oxidase, impregnated in a 0.004 inch thick nonwoven PET (e.g., Remay™ #2250). This composite
5 begins as roll stock from which circular discs are steel-rule die cut.

A plowfold liner having the general shape and dimensions shown in Figure 9E is produced from 0.0016
10 inch thick biaxially oriented polypropylene film coated on one side with silicone release (e.g., Fox River, #1803; Fox River Associates). This film is folded and perforated to length on a rotary press. The folded film is pulled apart at its perforations to create single
15 liners and is laminated to the outer adhesive surface of the retaining layer.

A patient liner having the general shape and dimensions shown in Figure 9F is produced (steel-rule die cut from sheet stock) from, for example, 0.003 inch
20 Fox River (#1806) PET coated on one side (facing/contacting the mask) with silicone release.

The components are assembled in the following order from the bottom up (for example, using manual assembly and human visual alignment): (1) sensor-to-tray
25 assembly; (2) plowfold liner; (3) gel disks (collection inserts); (4) mask layer; and (5) patient liner.

Figure 9G shows a plan view of the assembly just described with all components showing.

30 Accordingly, novel laminates, collection assemblies, and autosensor assemblies are disclosed. Although preferred embodiments of the subject invention

have been described in some detail, it is understood that obvious variations can be made without departing from the spirit and the scope of the invention as defined by the appended claims.

What is claimed is:

1. A collection assembly, for use in a
iontophoretic sampling device useful to monitor a
5 selected analyte or derivatives thereof present in a
biological system, said collection assembly comprising
 - a) a collection insert layer comprised of an
ionically conductive material having first and second
portions, each portion having first and second surfaces,
 - 10 b) a mask layer comprised of a material that is
substantially impermeable to the selected analyte or
derivatives thereof, wherein the mask layer (i) has
inner and outer faces and said outer face provides
contact with said biological system and the inner face
15 is positioned in facing relation with the first surface
of each collection insert, (ii) defines first and second
openings that are aligned with the first and second
portions of the collection insert layer, (iii) each
opening exposes at least a portion of the first surface
20 of the collection insert layer, and (iv) has a border
which extends beyond the first surface of each portion
of the collection insert layer to provide an overhang;
and
 - (c) a retaining layer having (i) inner and outer
25 faces wherein the inner face is positioned in facing
relation with the second surface of each collection
insert, (ii) defines first and second openings that are
aligned with the first and second portions of the
collection insert layer, (iii) each opening exposes at
30 least a portion of the second surface of the collection
insert layer, and (iv) has a border which extends beyond

the first surface of each portion of the collection insert layer to provide an overhang.

2. The collection assembly of claim 1, wherein (i)
5 the inner face of the mask layer contacts the inner face of the retaining layer, and (ii) the overhangs provided by the mask and retaining layers sandwich the collection insert layer portions therebetween.

10 3. The collection assembly of claim 2, wherein the perimeter of the mask layer is greater than the perimeter of the retaining layer thus forming an overhang of the mask layer relative to the perimeter of the retaining layer.

15

4. The collection assembly of claim 1, wherein said collection insert layer further comprises a gasket layer and the gasket layer is between the mask layer and the retaining layer.

20

5. The collection assembly of claim 4, wherein (i)
said gasket layer has first and second parts, each part comprising a substantially planar material having a top face, a bottom face, and an opening extending between
25 said top and bottom faces, wherein the top face of each gasket layer part contacts the inner face of the mask layer, the bottom face of each gasket layer part contacts the inner face of the retaining layer, and the openings of the first and second parts are each axially
30 aligned with the first and second openings in the mask and retaining layers, and (ii) within each gasket layer

part a collection insert is arranged and substantially fills each opening in each gasket layer part.

6. The collection assembly of claim 5, wherein
5 each opening of the retaining layer exposes the entire second surface of each collection insert, thus creating a gasket.

7. The collection assembly of claim 1, wherein
10 the mask layer is comprised of a material selected from the group consisting of high density polyethylene (HDPE), low density polyethylene (LDPE), very low density polyethylene (VLDPE), polyethylene copolymers, thermoplastic elastomers, silicon elastomers,
15 polyurethane (PU), polypropylene (PP), (PET), nylon, flexible polyvinylchloride (PVC), natural rubber, synthetic rubber, and combinations thereof.

8. The collection assembly of claim 3, wherein
20 the retaining layer is comprised of a material selected from the group consisting of high density polyethylene (HDPE), low density polyethylene (LDPE), very low density polyethylene (VLDPE), polyethylene copolymers, thermoplastic elastomers, silicon elastomers,
25 polyurethane (PU), polypropylene (PP), (PET), nylon, flexible polyvinylchloride (PVC), natural rubber, synthetic rubber, and combinations thereof.

9. The collection assembly of claim 3, wherein at
30 least one of the collection insert layer portions comprises glucose oxidase.

10. The collection assembly of claim 1, wherein the collection insert layer comprises a hydrogel.

11. The collection assembly of claim 1, wherein
5 the retaining layer has an outer face that is adhesive.

12. The collection assembly of claim 1, wherein the outer face of the retaining layer and exposed surfaces of the collection layer insert contact a first
10 surface of a first removable liner.

13. The collection assembly of claim 12, wherein said first removable liner has a plow-fold shape.

14. The collection assembly of claim 1, wherein
15 the outer face of the mask layer and exposed surfaces of the collection layer insert contact a first surface of a second removable liner.

15. The collection assembly of claim 1, further
20 comprising a first removable liner attached to the outer face of the retaining layer, and a second removable liner attached to the outer face of the mask layer.

16. The collection assembly of claim 1, wherein
25 the mask layer has an outer face that is adhesive.

17. The collection assembly of claim 1, wherein the retaining layer has an outer face that is adhesive.
30

18. The collection assembly of claim 1, wherein the first and second openings in the mask layer are

positioned in the collection assembly such that they are aligned with the first and second openings in the retaining layer and thereby define a plurality of flow paths through the collection assembly.

5

19. The collection assembly of claim 1, wherein the mask and retaining layers are contacted with each other along a central portion which separates the first and second openings in each layer such that said first and second portions of the collection insert are
10 individually sandwiched between the mask and retaining layers.

20. The collection assembly of claim 1, where said
15 analyte is glucose.

21. A laminate comprising any one of the collection assemblies of any of claims 1 through and including 20.

20

22. A sealed package containing the laminate of claim 21.

23. The sealed package of claim 22, further
25 comprising a hydrating insert.

24. An autosensor assembly for use in a iontophoretic sampling device useful to monitor an analyte present in a biological system, said autosensor assembly comprising,
30

(I) a collection assembly said collection assembly comprising,

a) a collection insert layer comprised of an ionically conductive material having first and second portions, each portion having first and second surfaces,

b) a mask layer comprised of a substantially planar material that is substantially impermeable to the selected analyte or derivatives thereof, wherein the mask layer (i) has inner and outer faces and said outer face provides contact with said biological system and the inner face is positioned in facing relation with the first surface of each collection insert, (ii) defines first and second openings that are aligned with the first and second portions of the collection insert layer, (iii) each opening exposes at least a portion of the first surface of the collection insert layer, and (iv) has a border which extends beyond the first surface of each portion of the collection insert layer to provide an overhang;

(c) a retaining layer having (i) inner and outer faces wherein the inner face is positioned in facing relation with the second surface of each collection insert, (ii) defines first and second openings that are aligned with the first and second portions of the collection insert layer, (iii) each opening exposes at least a portion of the second surface of the collection insert layer, and (iv) has a border which extends beyond the first surface of each portion of the collection insert layer to provide an overhang; and

(d) where the first and second openings in the mask layer are positioned in the collection assembly such that they are aligned with the first and second openings in the retaining layer and thereby define a plurality of flow paths through said collection assembly;

(II) an electrode assembly having an inner and outer face, the inner face comprising first and second bimodal electrodes, wherein the first and second bimodal electrodes are aligned with the first and second openings in the retaining layer of the collection assembly; and

(III) a support tray that contacts the outer face of the electrode assembly.

10 25. The autosensor assembly of claim 24, wherein the collection assembly (i) the inner face of the mask layer contacts the inner face of the retaining layer, and (ii) the overhangs provided by the mask and retaining layers sandwich the collection insert layer portions therebetween.

26. The autosensor assembly of claim 25, wherein the perimeter of the mask layer is greater than the perimeter of the retaining layer.

20

27. The autosensor assembly of claim 24, wherein the mask layer is comprised of a material selected from the group consisting of high density polyethylene (HDPE), low density polyethylene (LDPE), very low density polyethylene (VLDPE), polyethylene copolymers, thermoplastic elastomers, silicon elastomers, polyurethane (PU), polypropylene (PP), (PET), nylon, flexible polyvinylchloride (PVC), natural rubber, synthetic rubber, and combinations thereof.

30

28. The autosensor assembly of claim 24, wherein the retaining layer is comprised of a material selected

from the group consisting of high density polyethylene (HDPE), low density polyethylene (LDPE), very low density polyethylene (VLDPE), polyethylene copolymers, thermoplastic elastomers, silicon elastomers,
5 polyurethane (PU), polypropylene (PP), (PET), nylon, flexible polyvinylchloride (PVC), natural rubber, synthetic rubber, and combinations thereof.

29. The autosensor assembly of claim 24, wherein
10 at least one of the collection insert layer portions comprises glucose oxidase.

30. The autosensor assembly of claim 24, wherein the collection insert layer comprises a hydrogel.
15

31. The autosensor assembly of claim 24, wherein the retaining layer has an outer face that is adhesive.

32. The autosensor assembly of claim 24, wherein
20 the outer face of the retaining layer and exposed surfaces of the collection layer insert contact a first surface of a first removable liner.

33. The autosensor assembly of claim 32, wherein
25 said first removable liner has a plow-fold shape.

34. The autosensor assembly of claim 24, wherein the outer face of the mask layer and exposed surfaces of the collection layer insert contact a first surface of a
30 second removable liner.

35. The autosensor assembly of claim 24, further comprising a first removable liner attached to the outer face of the retaining layer, and a second removable liner attached to the outer face of the mask layer.

5

36. The autosensor assembly of claim 24, wherein the mask layer has an outer face that is adhesive.

37. The autosensor assembly of claim 24, wherein
10 the retaining layer has an outer face that is adhesive.

38. The autosensor assembly of claim 24, wherein the mask and retaining layers are contacted with each other along a central portion which separates the first
15 and second openings in each layer such that said first and second portions of the collection insert are individually sandwiched between the mask and retaining layers.

20 39. The autosensor assembly of claim 24, wherein said analyte is glucose.

40. The autosensor assembly of any of claims 24 through and including 39, wherein said collection
25 assembly is a laminate.

41. A sealed package containing the autosensor assembly of claim 40.

30 42. The sealed package of claim 41, further comprising a hydrating insert.

43. A sealed package containing the autosensor assembly of any of claims 24 through and including 39.

44. The sealed package of claim 43, further comprising a hydrating insert.

45. A collection assembly for use in a iontophoretic sampling device useful to monitor a selected analyte, or derivatives thereof, present in a biological system, said collection assembly comprising

a) a mask layer comprised of a substantially planar material that is substantially impermeable to the selected analyte or derivatives thereof, where said mask layer has inner and outer faces and said outer face provides contact with said biological system;

b) a collection insert layer comprised of an ionically conductive material having first and second surfaces, and

c) said mask and collection insert layers are configured such that (i) at least a portion of said collection insert is exposed to provide contact with the biological system, and (ii) flow of the analyte through the first surface of the collection insert layer from the biological system is prevented by the mask layer for any portion of the first surface of the collection insert layer that is in contact with the inner face of the mask layer.

46. An autosensor assembly comprising

(a) the collection assembly of claim 45,

(b) an electrode assembly having an inner face comprising an electrode and an outer face, where the inner face of the electrode assembly and the collection assembly are aligned to define a plurality of flow paths
5 through said collection assembly, and

(c) a support tray that contacts said outer face of the electrode assembly.

47. A sealed package containing the autosensor
10 assembly of claim 46.

48. The sealed package of claim 47, further comprising a hydrating insert.

15

1/16

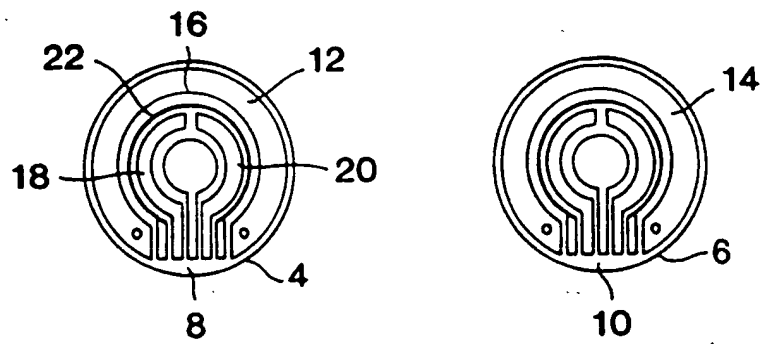


FIG. 1A

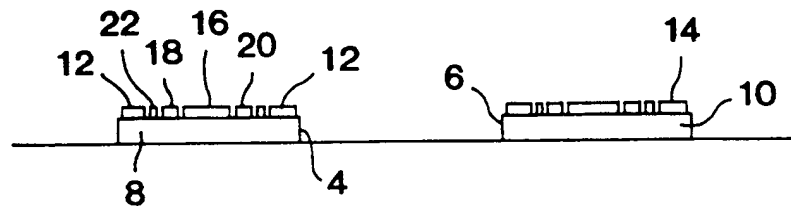


FIG. 1B

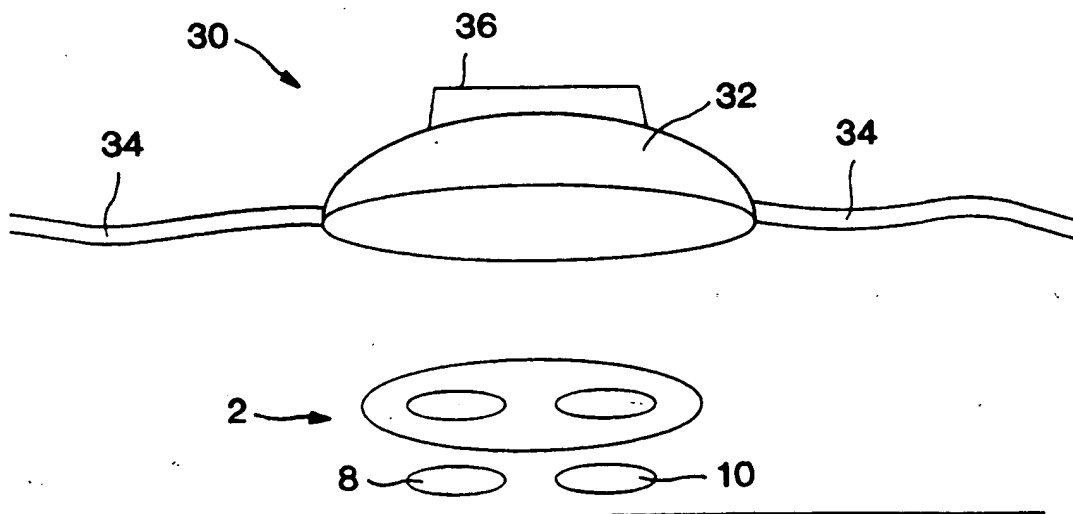


FIG. 2

2/16

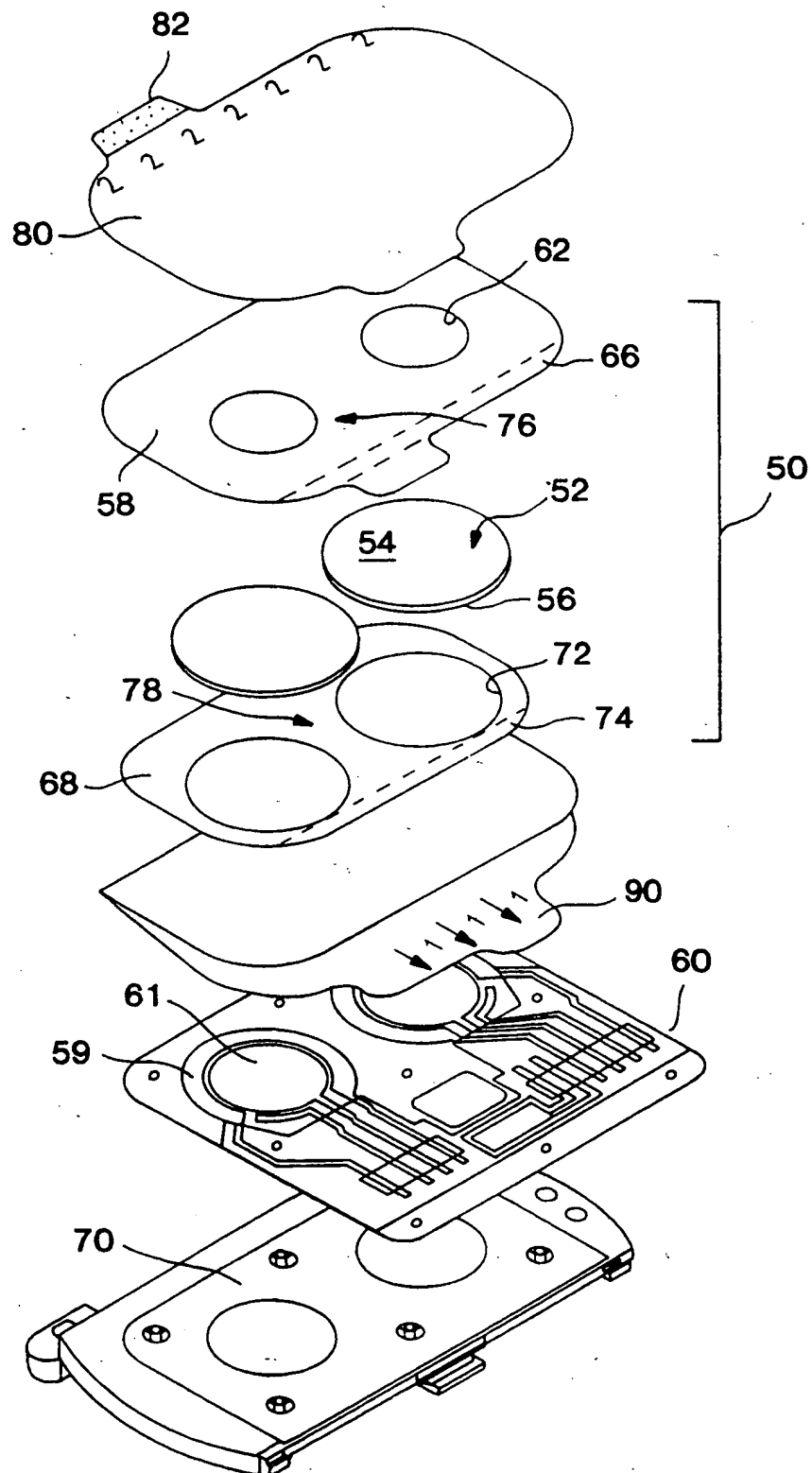


FIG. 3

3/16

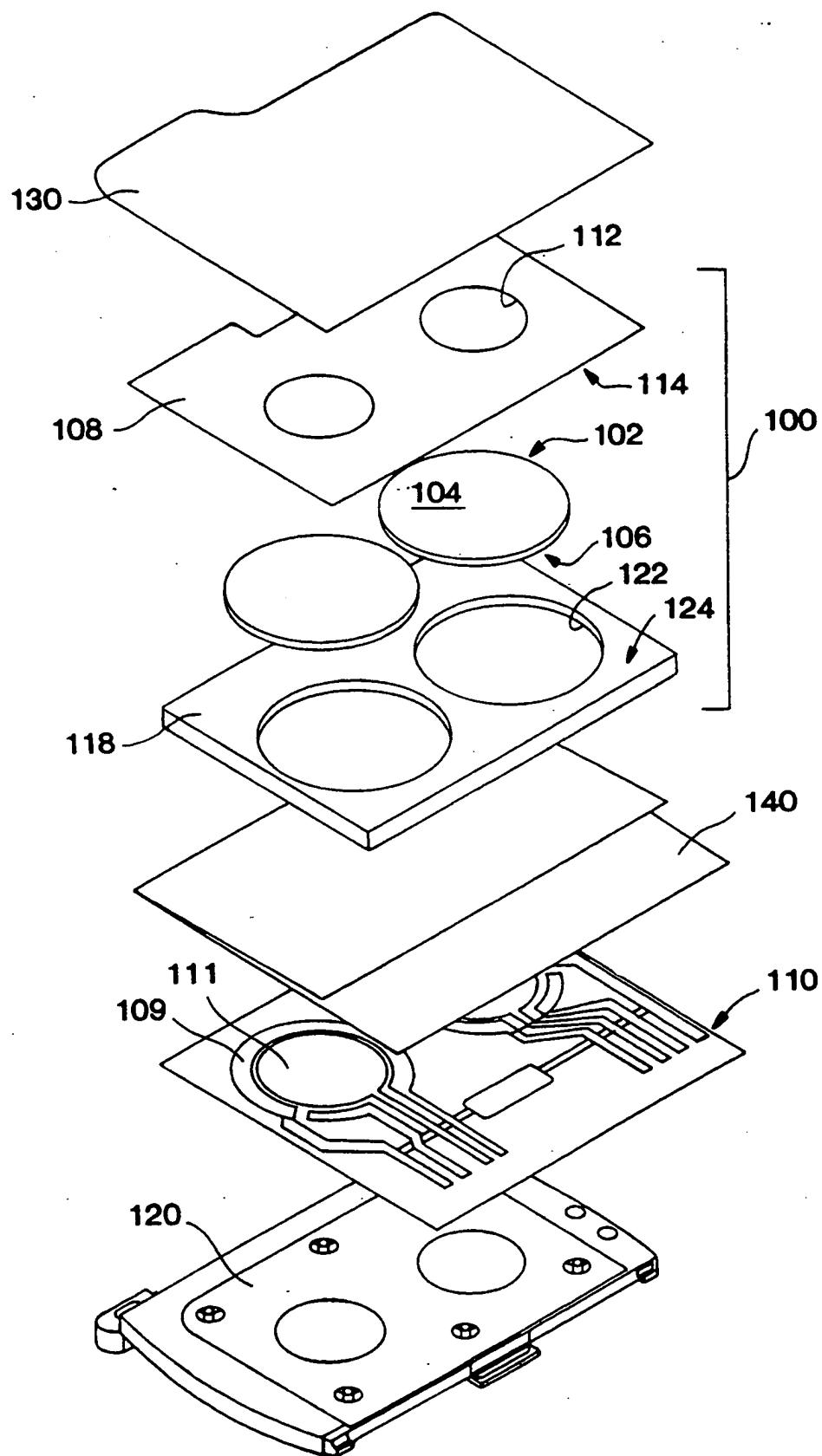


FIG. 4

4/16

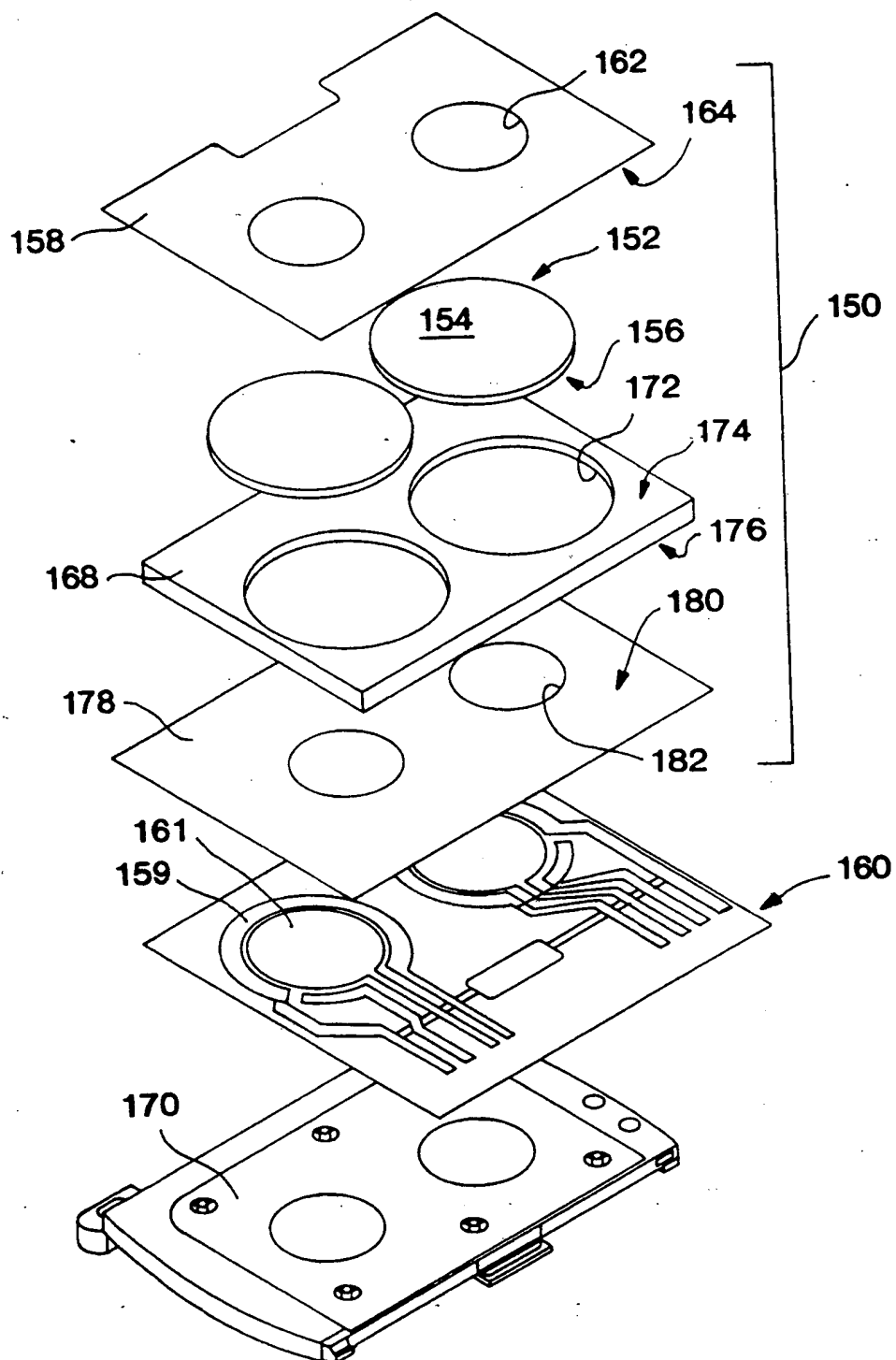


FIG. 5

5/16

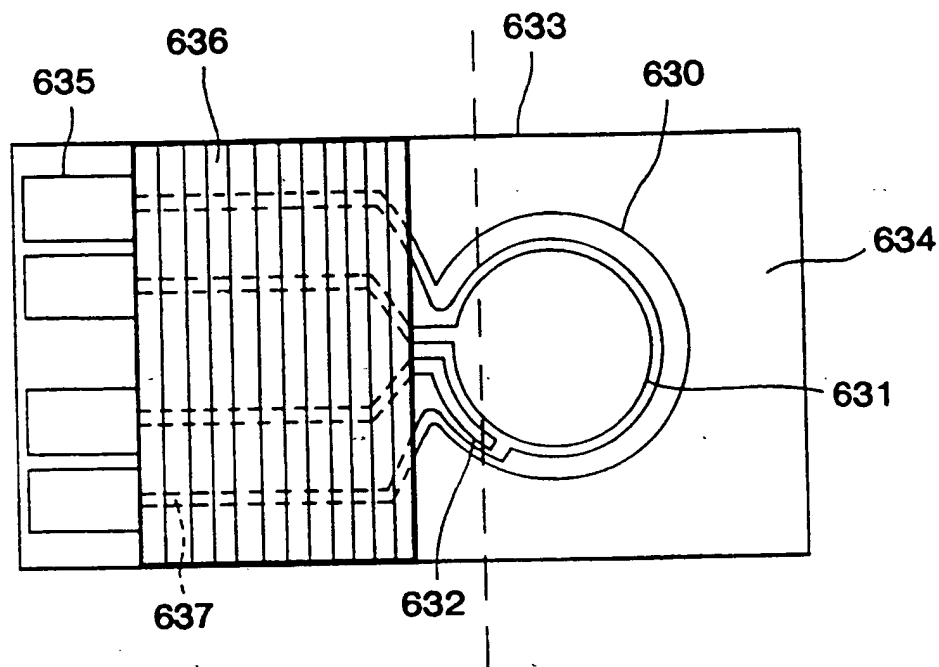


FIG. 6

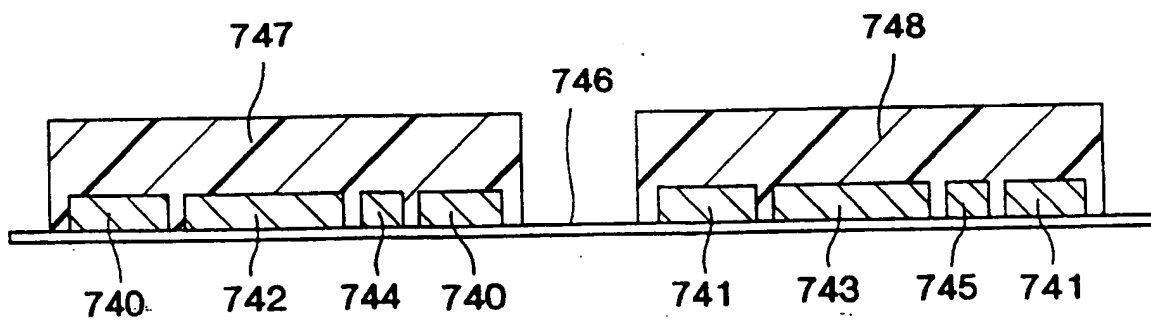


FIG. 7

6/16

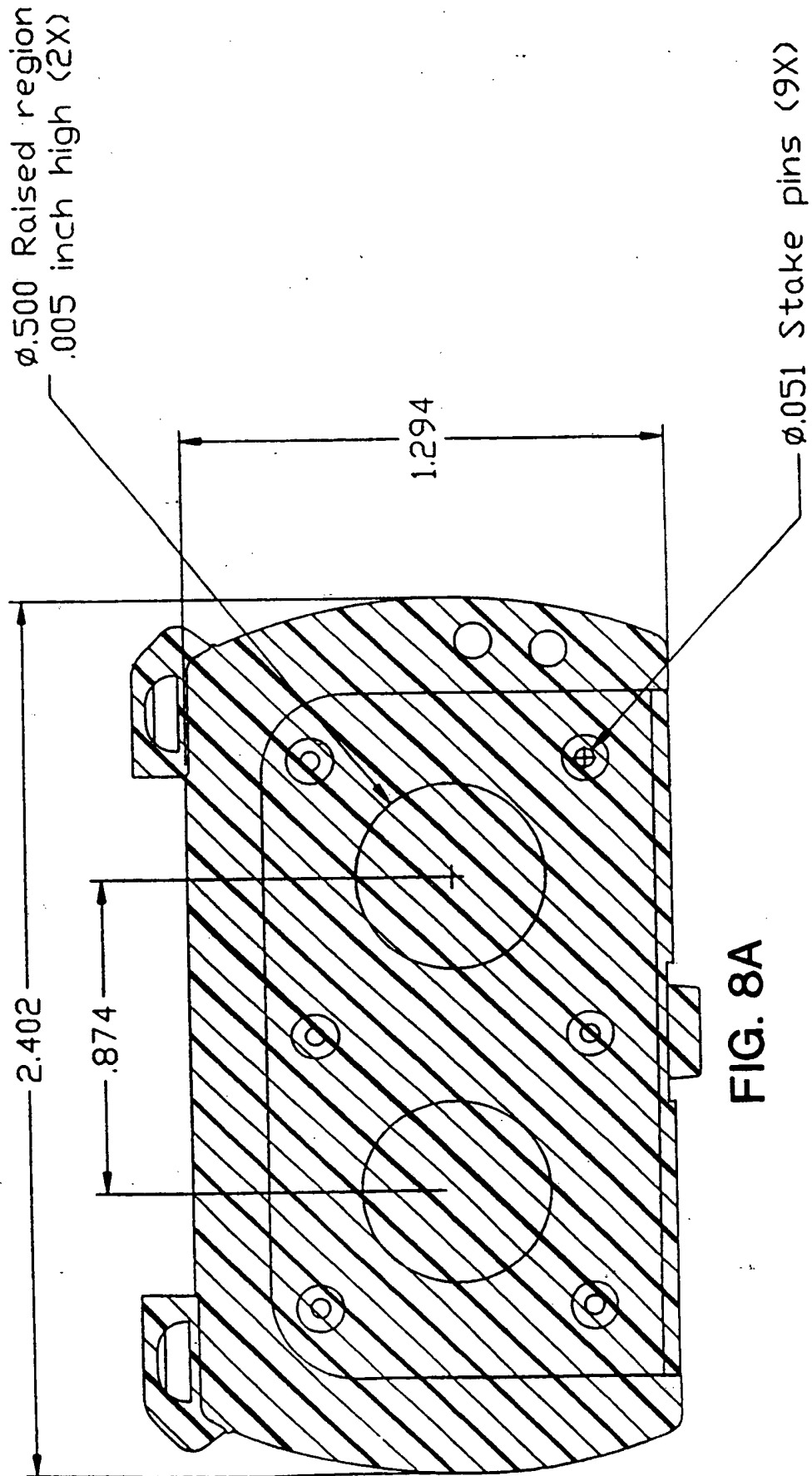


FIG. 8A

7/16

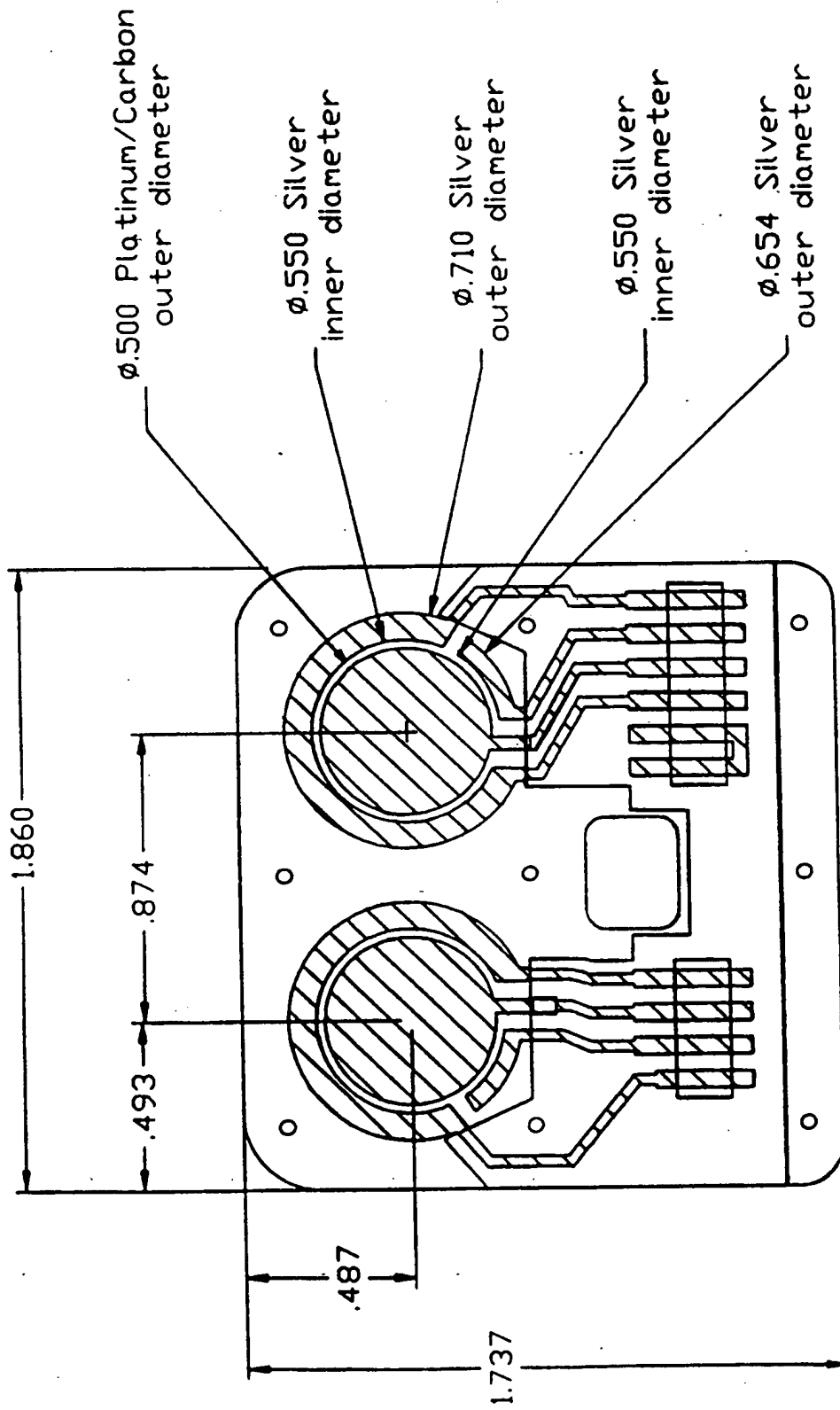


FIG. 8B

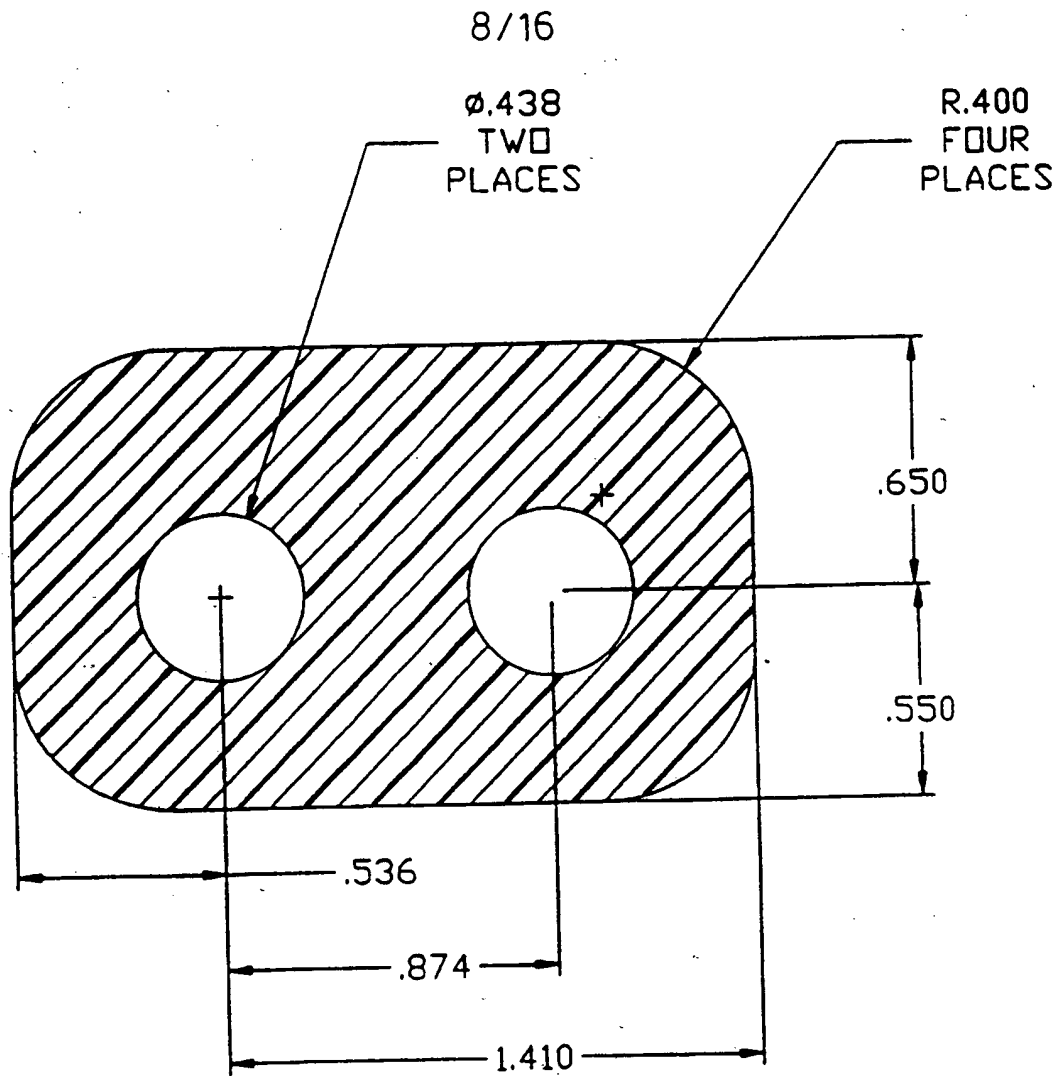


FIG. 8C

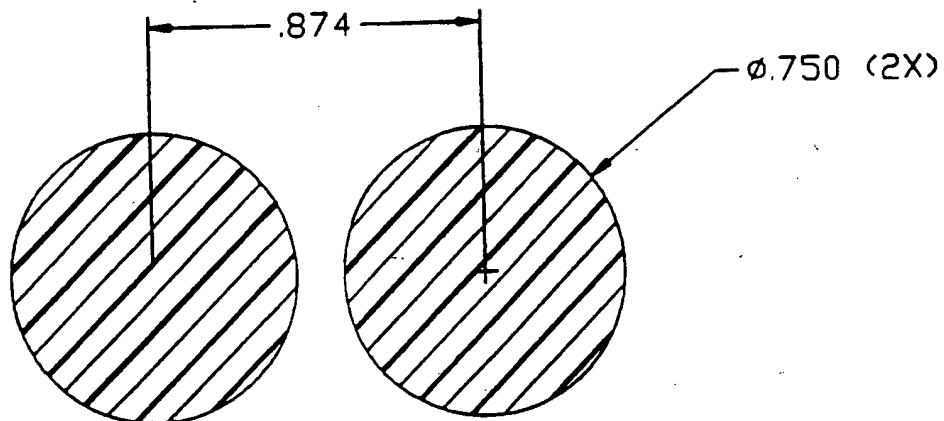


FIG. 8D

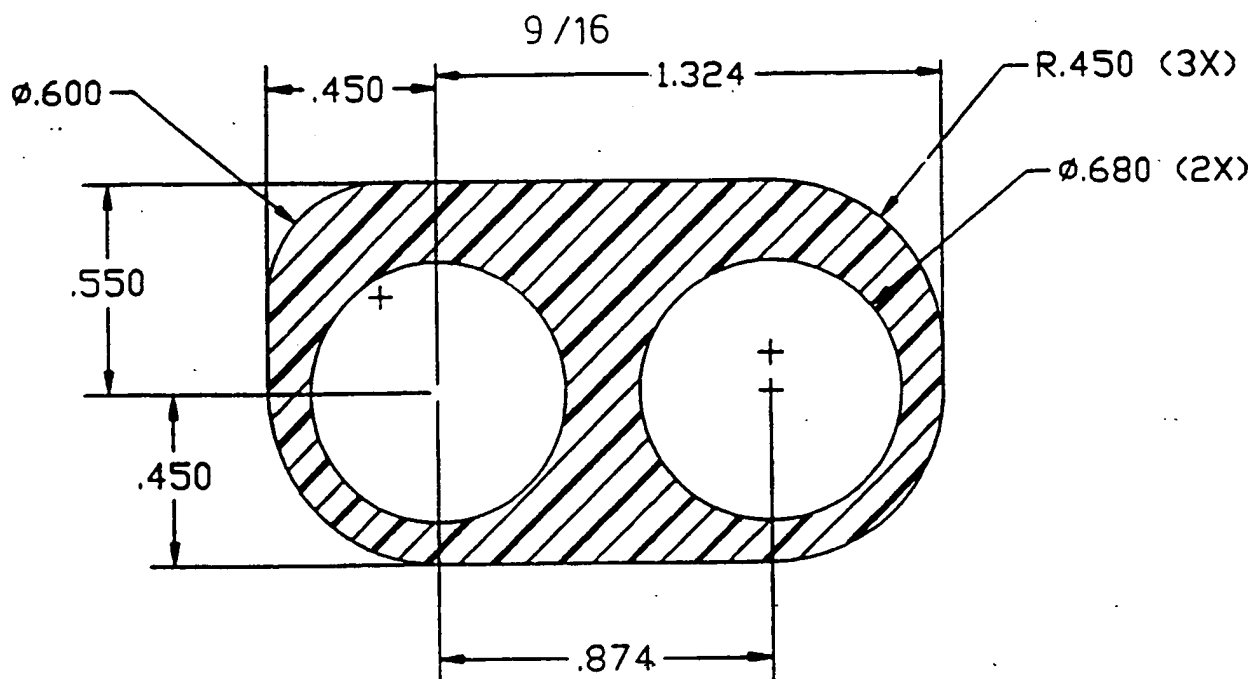


FIG. 8E

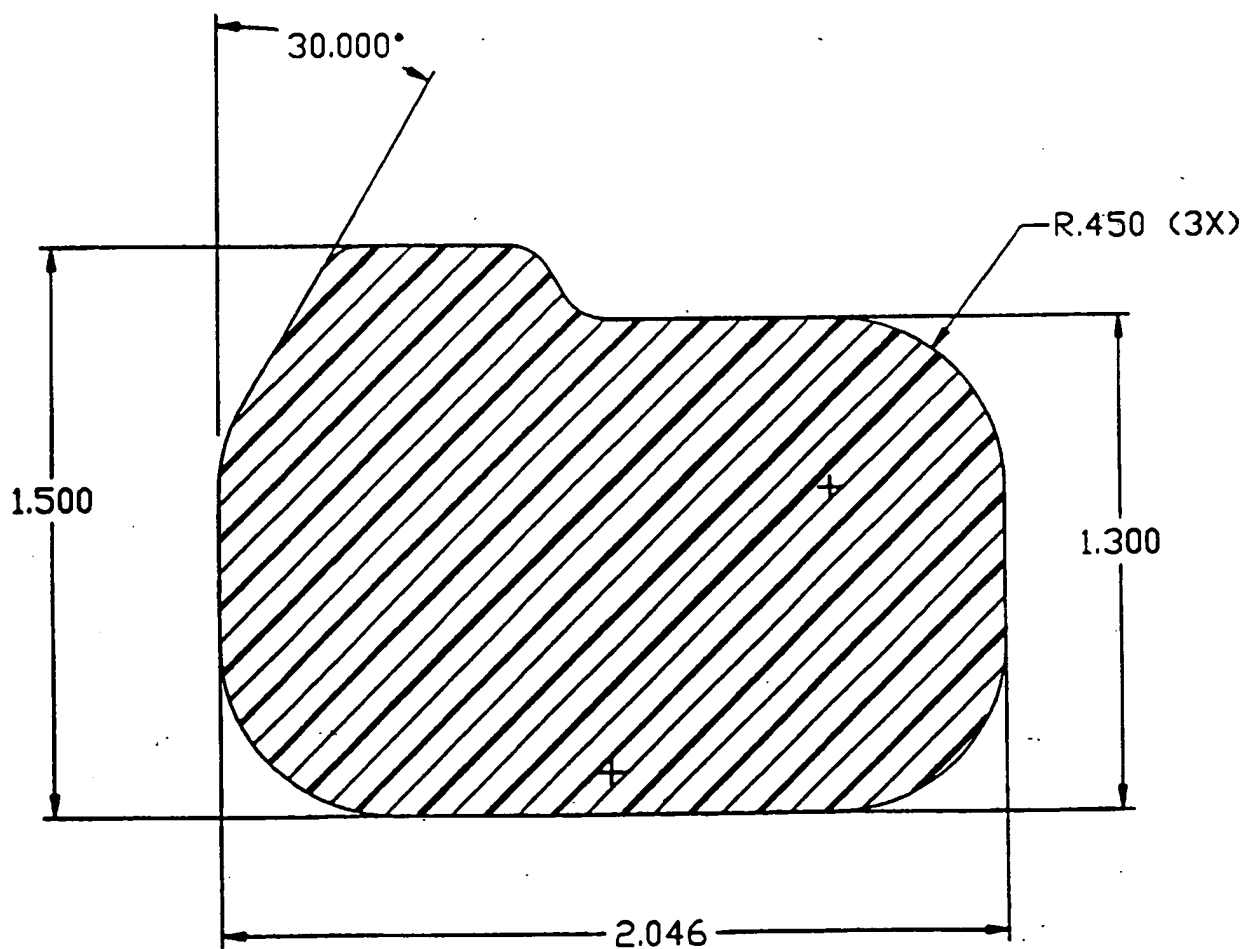


FIG. 8F

10/16

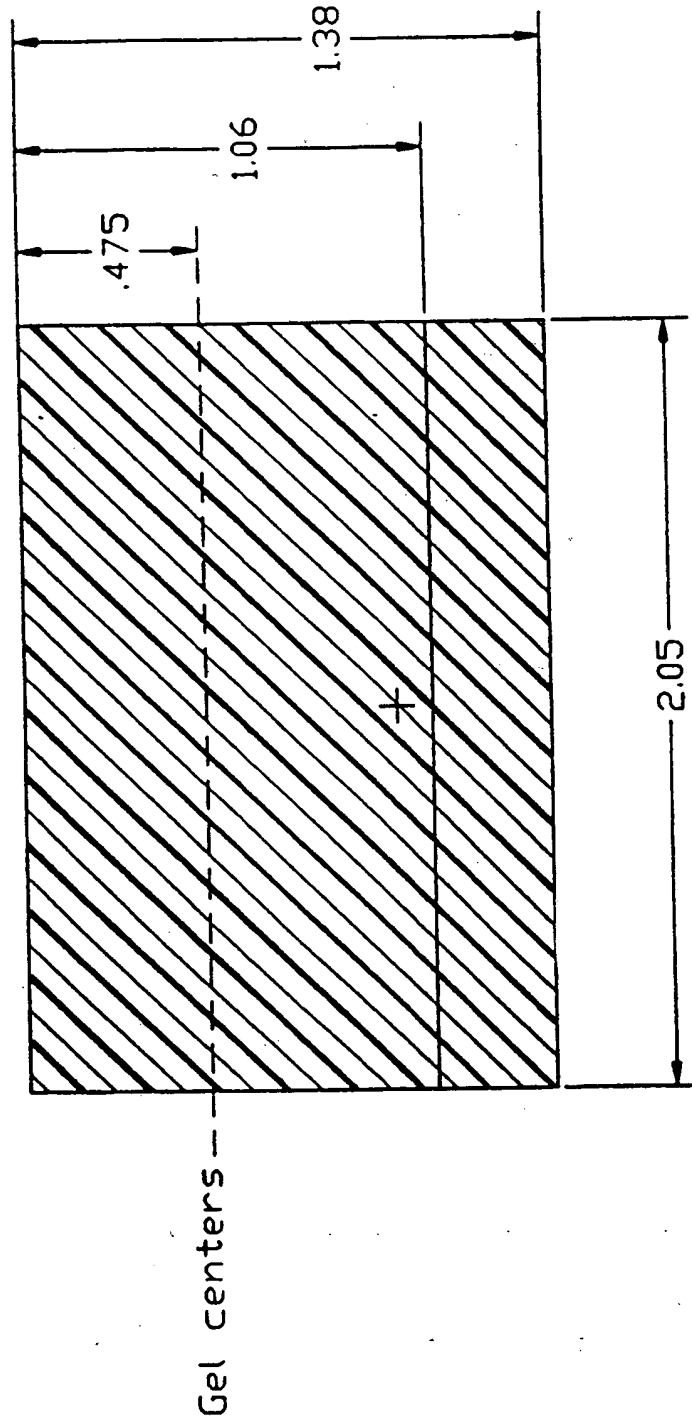


FIG. 8G

11/16

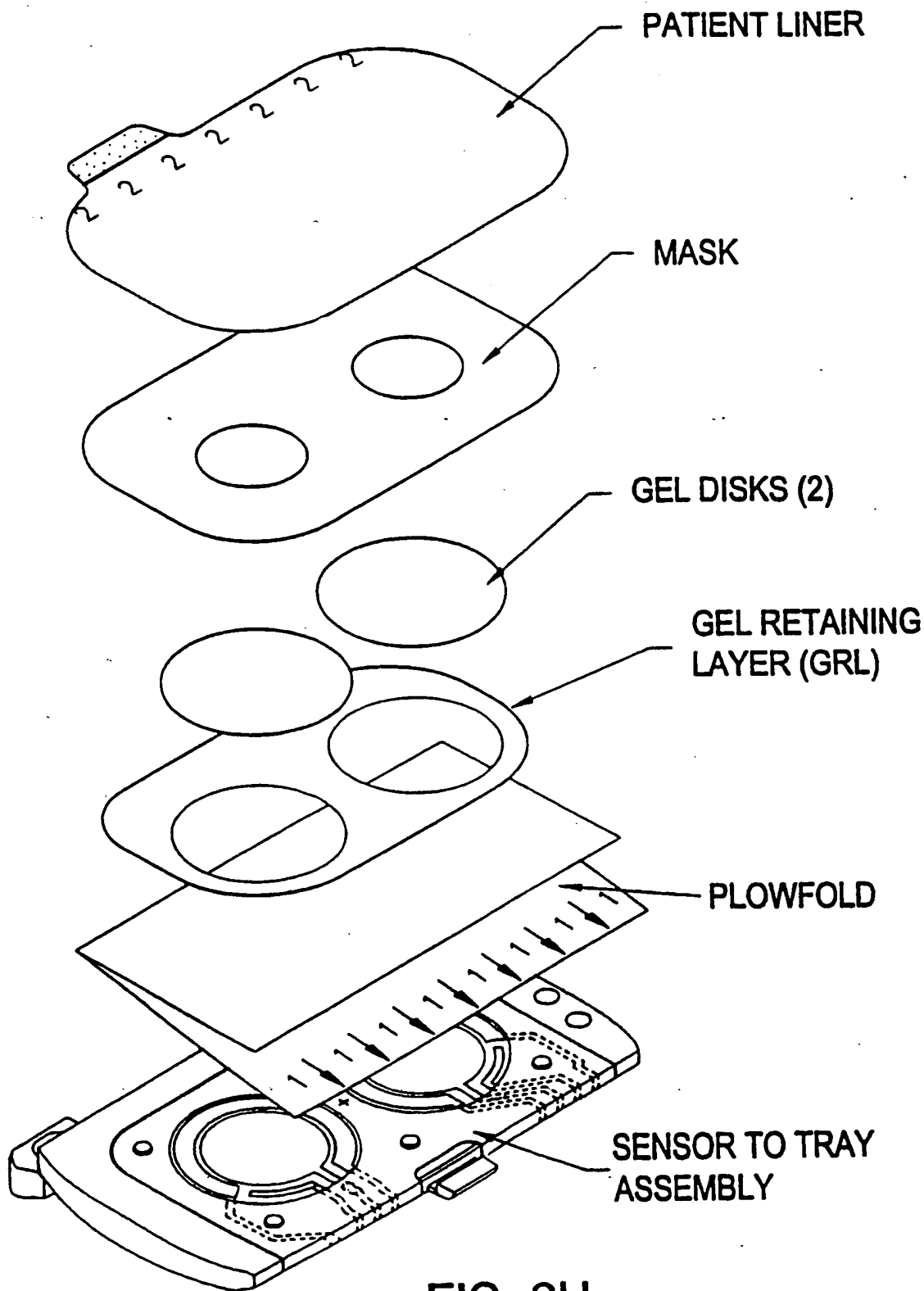


FIG. 8H

12 / 16

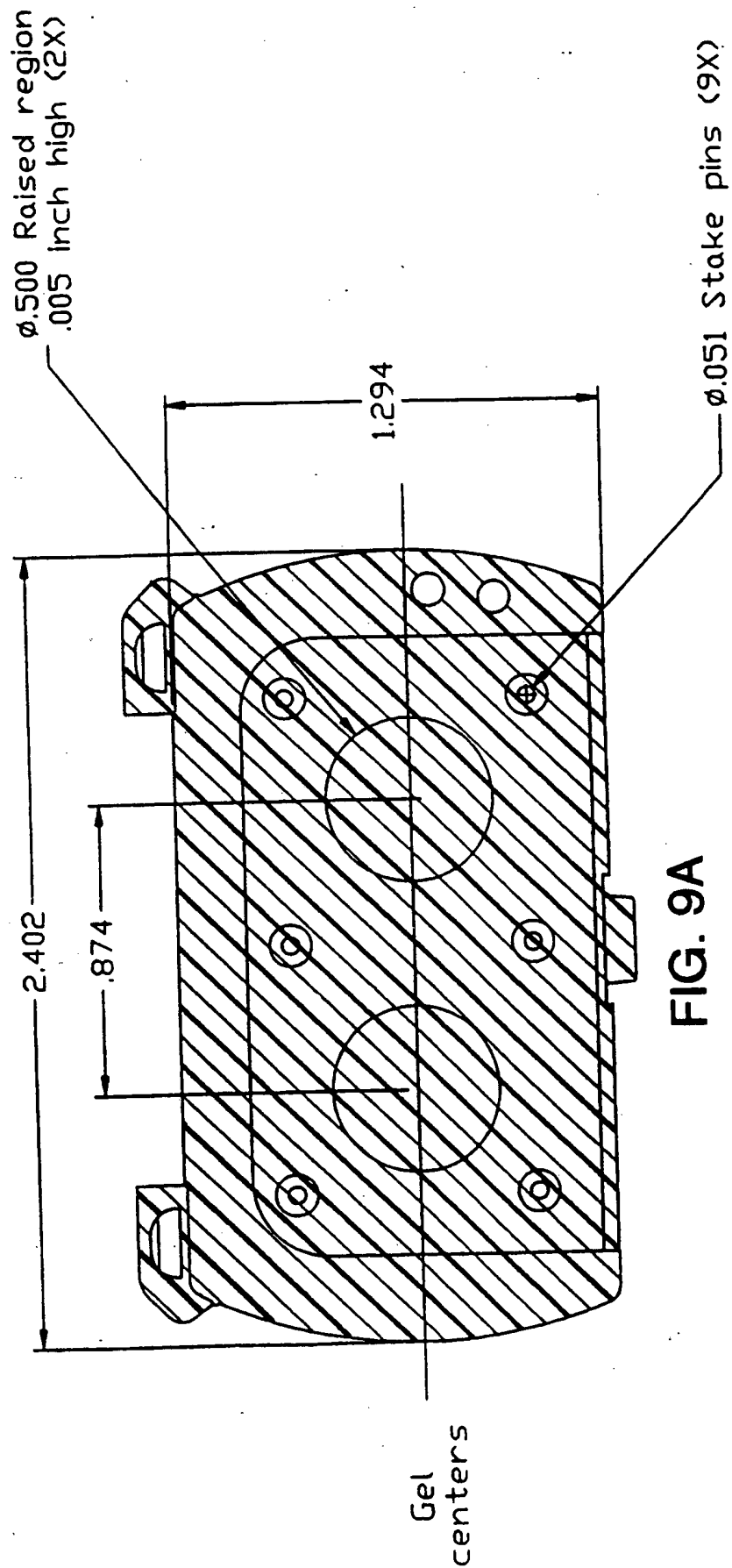


FIG. 9A



14/16

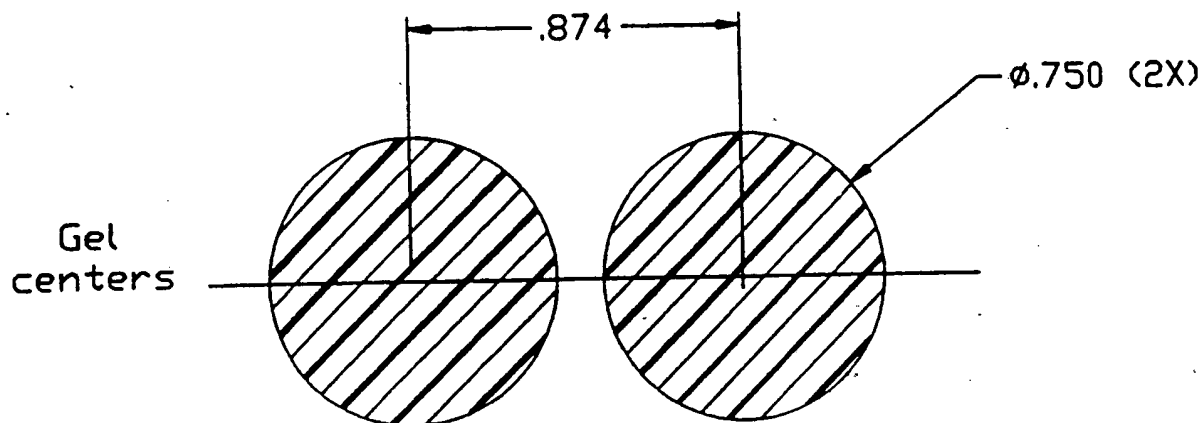


FIG. 9D

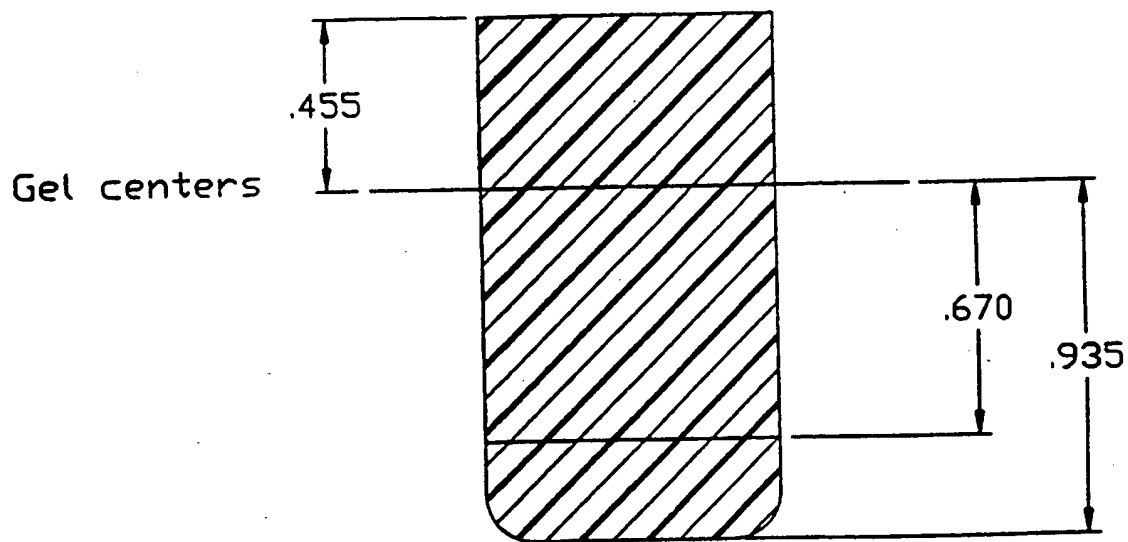


FIG. 9E

15/16

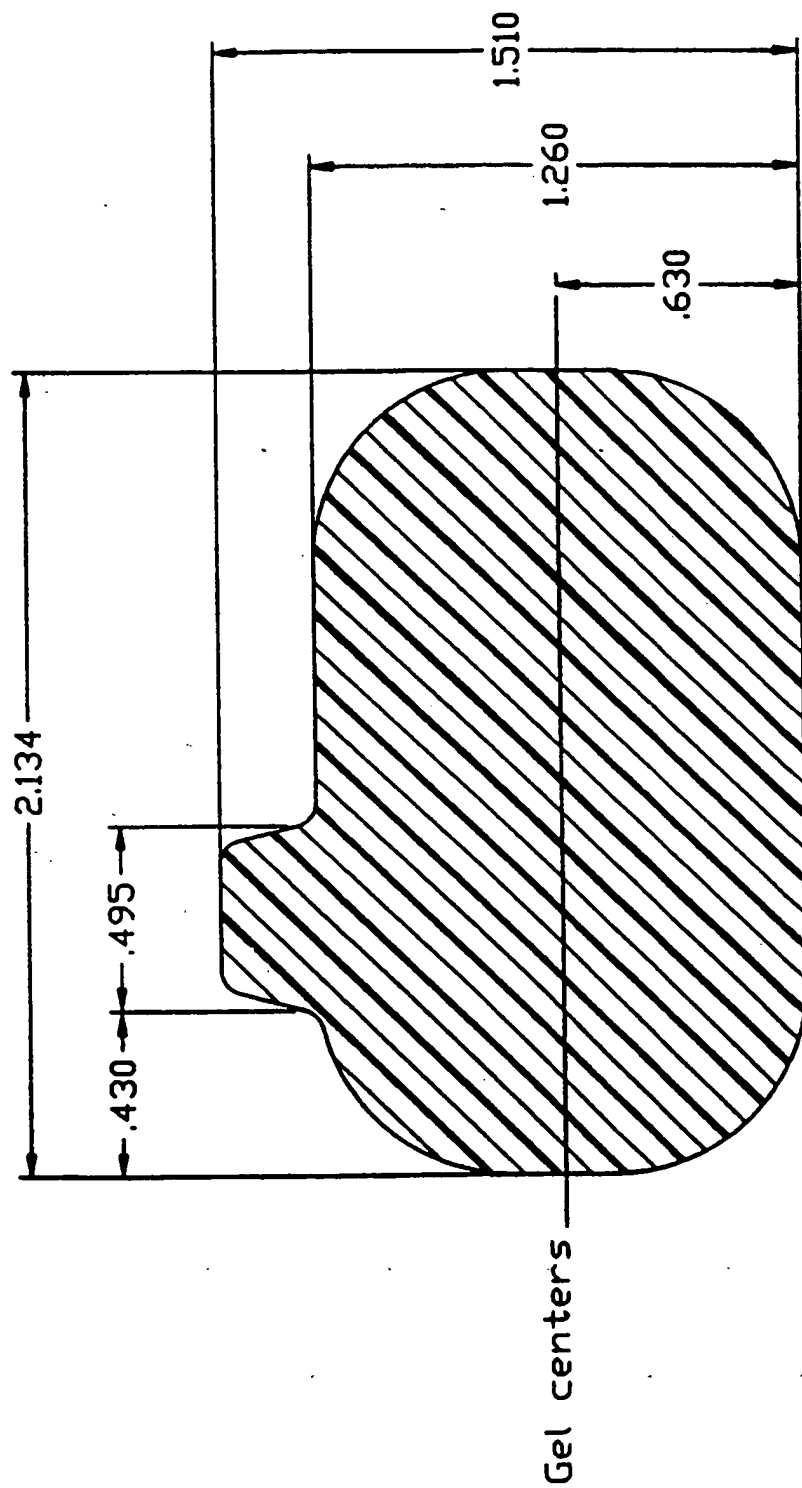


FIG. 9F

16/16

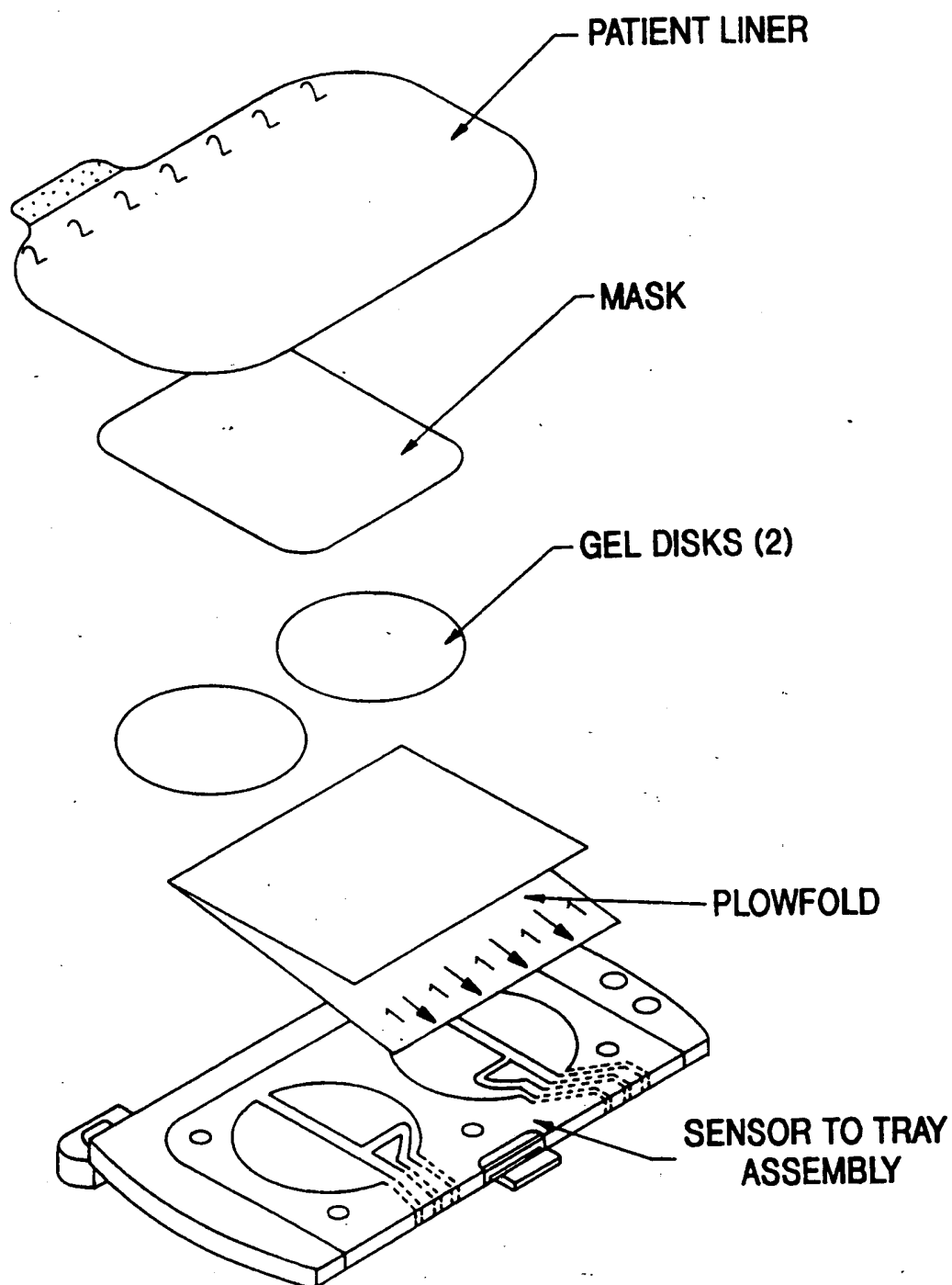


FIG. 9G

A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 A61N1/30

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 A61N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO 96 00110 A (CYGNUS THERAPEUTIC SYSTEMS) 4 January 1996 (1996-01-04) page 18, line 3 - page 21, line 5 page 24, line 26 - page 27, line 21; figures 1A-1D, 37-39 ---	1-4, 8-48
A	WO 97 10356 A (CYGNUS THERAPEUTIC SYSTEMS) 20 March 1997 (1997-03-20) page 18, line 29 - page 24, line 23 page 26, line 32 - page 29, line 28; figures 1-6 --- -/--	1-3, 7-12, 16-32, 39-46



Further documents are listed in the continuation of box C.



Patent family members are listed in annex.

* Special categories of cited documents:

- "A" document defining the general state of the art which is not considered to be of particular relevance
- "E" earlier document but published on or after the international filing date
- "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the international filing date but later than the priority date claimed

- "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- "&" document member of the same patent family

Date of the actual completion of the international search

10 September 1999

Date of mailing of the international search report

21/09/1999

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2
NL - 2280 HV Rijswijk
Tel. (+31-70) 340-2040, Tx. 31 651 epo nl,
Fax: (+31-70) 340-3016

Authorized officer

Rakotondrajaona, C

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	US 5 279 543 A (CULLANDER CHRISTOPHER ET AL) 18 January 1994 (1994-01-18) column 11, line 36 - column 15, line 46 column 17, line 40 - column 18, line 59; figures -----	1-3, 19-21, 24, 38-41,45
A	EP 0 625 360 A (TACHIBANA KATSURO ;TACHIBANA SHUNRO (JP)) 23 November 1994 (1994-11-23) page 4, column 5, line 22 - page 5, column 8, line 31; figures 1-3 -----	1,45
A	WO 89 06555 A (MASSACHUSETTS INST TECHNOLOGY) 27 July 1989 (1989-07-27) page 7, line 29 - page 9, line 19; figures -----	1,45

Patent document cited in search report		Publication date	Patent family member(s)	Publication date
WO 9600110	A	04-01-1996	AU 2944995	19-01-1996
			CA 2193885	04-01-1996
			EP 0766578	09-04-1997
			JP 10506293	23-06-1998
WO 9710356	A	20-03-1997	US 5735273	07-04-1998
			AU 703849	01-04-1999
			AU 6497396	01-04-1997
			CA 2229509	20-03-1997
			EP 0876501	11-11-1998
			US 5827183	27-10-1998
US 5279543	A	18-01-1994	AT 129909	15-11-1995
			AU 639888	12-08-1993
			AU 3183889	25-08-1989
			DE 68924716	14-12-1995
			DE 68924716	04-07-1996
			DK 179390	28-07-1990
			EP 0326398	02-08-1989
			EP 0673622	27-09-1995
			ES 2085863	16-06-1996
			IE 63406	19-04-1995
			JP 4502561	14-05-1992
			KR 9711449	11-07-1997
			PT 89560	10-11-1989
			WO 8906989	10-08-1989
			US 5730714	24-03-1998
			US 5362307	08-11-1994
EP 0625360	A	23-11-1994	JP 6070987	15-03-1994
			DE 69325572	12-08-1999
			US 5582586	10-12-1996
			WO 9405368	17-03-1994
WO 8906555	A	27-07-1989	AT 131081	15-12-1995
			CA 1326268	18-01-1994
			DE 68925030	18-01-1996
			DE 68925030	25-07-1996
			EP 0398960	28-11-1990
			JP 2798459	17-09-1998
			JP 3502416	06-06-1991
			US 5547467	20-08-1996
			US 5389069	14-02-1995
			US 5019034	28-05-1991
			US 5749847	12-05-1998
			US 5667491	16-09-1997

